

bacteria

cilia

nonciliated cells

Peter Sebo

Institute of Microbiology of the CAS, Prague

Baltimore 2010 / Dublin 2013 / Annecy 2015:

thanks to

‘**democracy**’

and

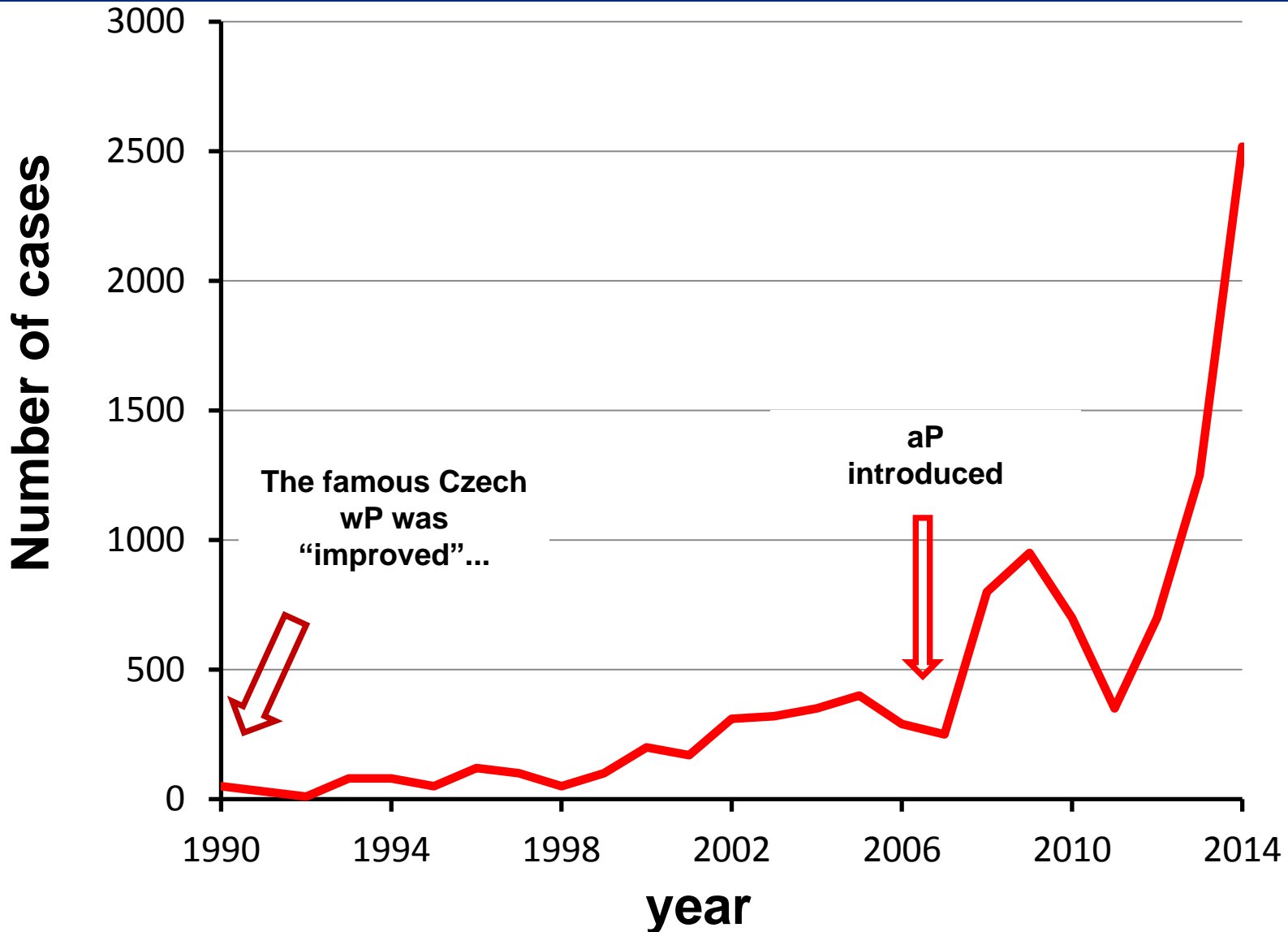
**acellular vaccines**

whooping cough is back

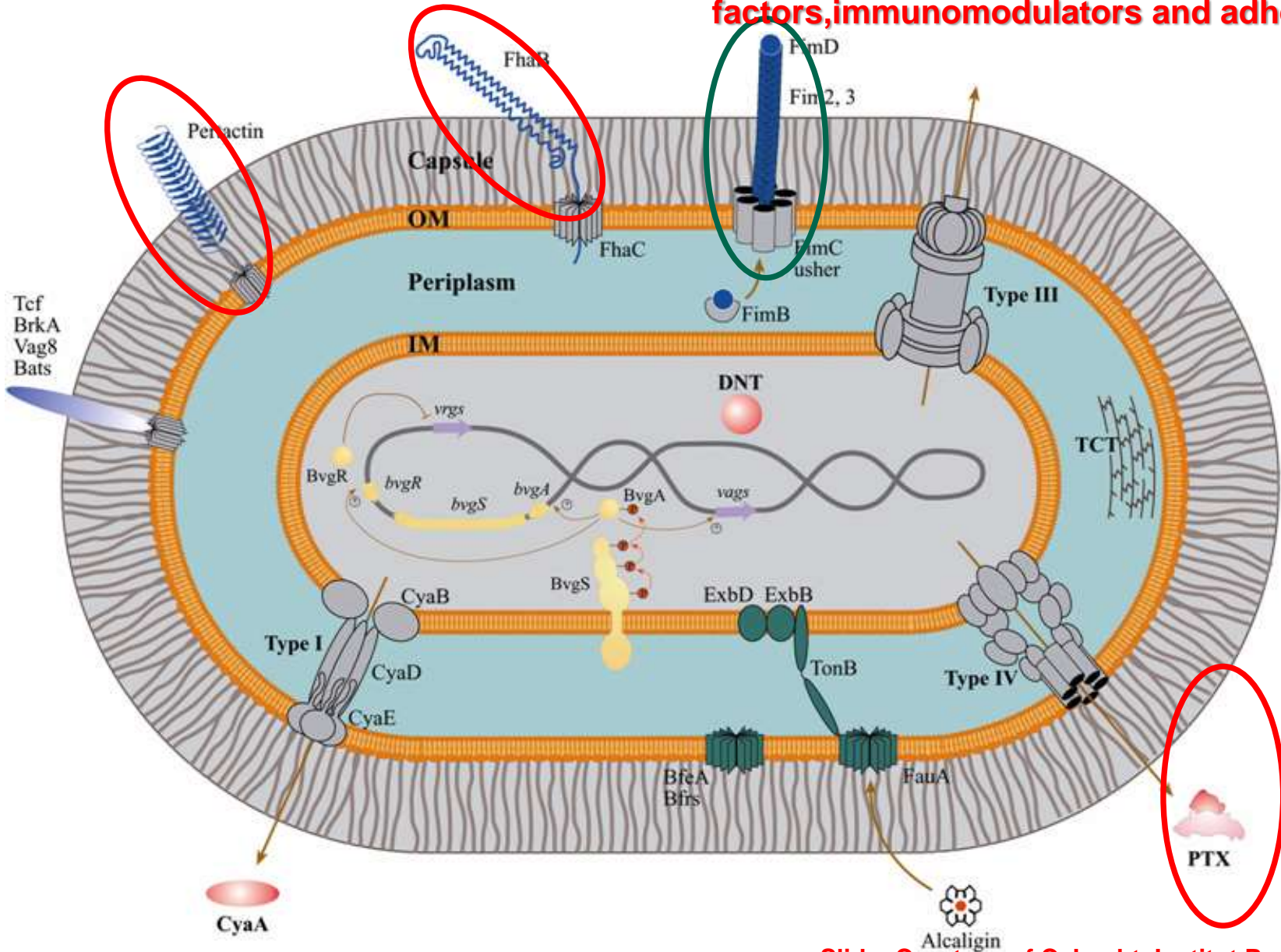
(to the wealthiest countries...)

# We've got a problem again...

Confirmed clinical pertussis in 1990-2013 in CR (10 million people)  
We had 2,518 cases in 2014 (like in 1961...)



***B. pertussis* is armed with numerous parallel and redundant virulence systems (cytotoxins, complement resistance factors, immunomodulators and adhesins)**



aP vaccines can be improved, but...

only a next generation of non-reactogenic wP vaccine will be able to stop the spread of the bug in the wealthiest populations

be it as a booster in older kids and adults – inducing sterilizing immunity or as priming in a pediatric hexavaccine

# ACT relevance for pertussis vaccines

# A bit of history - I.

- 1976 ACT discovered by Hewlett EL, Urban MA, Manclark CR, Wolff J.:  
'**Extracytoplasmic adenylate cyclase of *Bordetella pertussis***'. PNAS 73:1926-30.
- 1977 Hewlett EL, Manclark CR, Wolff J.: **Adenyl cyclase in *Bordetella pertussis* vaccines.**  
J Infect Dis. 1977 Aug;136 Suppl:S216-9
- 1980 Wolff *et al.*: **Calmodulin activates prokaryotic adenylate cyclase.** PNAS 77: 3841
- 1982 Confer DL and Eaton JW: **Phagocyte impotence caused by an invasive bacterial adenylate cyclase.** Science 217:948:

*...For unknown reasons, humans infected with the bacterium *Bordetella pertussis* are exceptionally vulnerable to secondary infections. *Bordetella* species elaborate a soluble, heat-stable, and highly active adenylate cyclase. This enzyme is internalized by phagocytic cells and catalyzes the unregulated formation of adenosine 3',5'-monophosphate (cyclic AMP), thereby disrupting normal cellular function. This unusual phenomenon may explain *Bordetella*-induced aphyllaxis...*

**aphyllaxis = absence of phyllaxis or immunity**

Obsolete term meaning lack of protection against disease

Lack of protection against disease. Also called *nonimmunity*.

Phagocyte impotence caused by an invasive bacterial adenylate cyclase. :

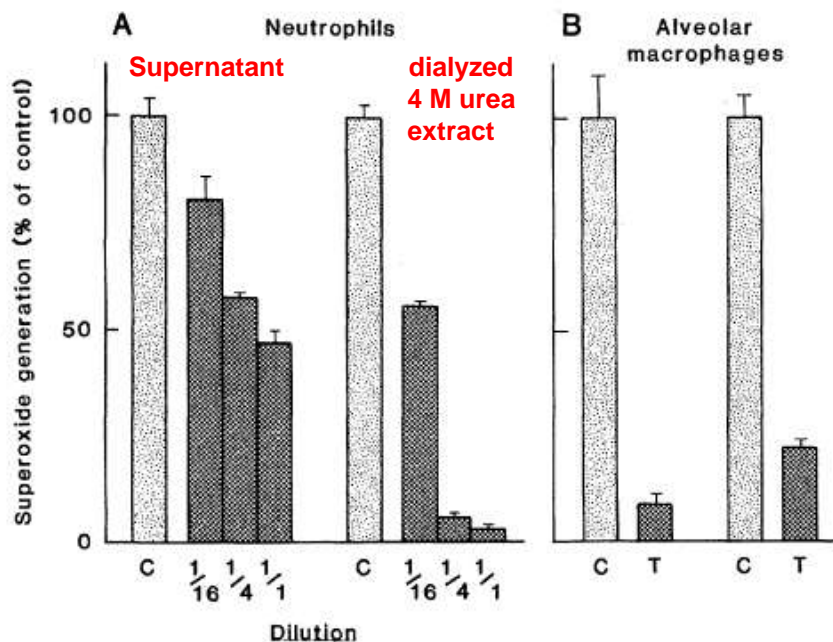


Fig. 1. Superoxide generation by stimulated human phagocytes and inhibition by *Bordetella* products. (A) Human neutrophils,  $2 \times 10^6$ , suspended in 200  $\mu$ l of Hanks balanced salt solution, were incubated for 10 minutes at 37°C with 200  $\mu$ l of the indicated dilution of the supernatant of 48-hour cultures of *B. pertussis* (protein content, 120  $\mu$ g/ml) (left panel) and with dialyzed extract of *B. pertussis* organisms (protein content, 520  $\mu$ g/ml) (right panel). Cytochrome *c* (1.2 mg) and opsonized zymosan (1 mg) were added (total volume, 1 ml) and the superoxide-dependent reduction of ferricytochrome *c* was determined after 10 additional minutes of incubation at 37°C as previously described (8). Results are expressed as percentages of control (untreated values), and bars represent the range of independent triplicate determinations. (B) Human alveolar macrophages ( $10^6$ ) suspended in 100  $\mu$ l of Hanks balanced salt solution were incubated with 100  $\mu$ l of dialyzed extract of *B. pertussis* (T) or 100  $\mu$ l of external dialysis fluid (C) as above. The cells were then stimulated by the addition of 1 mg of opsonized zymosan (left bars) or 0.1  $\mu$ g of phorbol myristate acetate (right bars). Superoxide production was assessed by following luminol-enhanced chemiluminescence as described (11). Results represent the mean and range of triplicate determinations.

**Culture supernatants contain very little of active ACT = huge potency!**

Sample	Treatment	Cyclic AMP (pmole/10 <sup>7</sup> PMN)	Adenylate cyclase (pmole/10 <sup>7</sup> PMN-min)
Neutrophils	Incubated for 20 minutes at 37°C, washed, trypsinized, washed, homogenized	4.9	0, 0*
Neutrophils plus <i>B. pertussis</i> extract (540 $\mu$ g/10 <sup>7</sup> PMN)	Incubated for 20 minutes at 37°C, washed, trypsinized, washed, homogenized	1296	41.9, 28.0, 45.1
Neutrophils plus <i>B. pertussis</i> extract (540 $\mu$ g/10 <sup>7</sup> PMN)	Incubated for 20 minutes at 0°C, washed, trypsinized, washed, homogenized	6.7	4.2, 4.3, 4.8

\*Limit of detection, < 1 pmole per 10<sup>7</sup> PMN per minute.



## Phagocyte impotence caused by an invasive bacterial adenylate cyclase. :

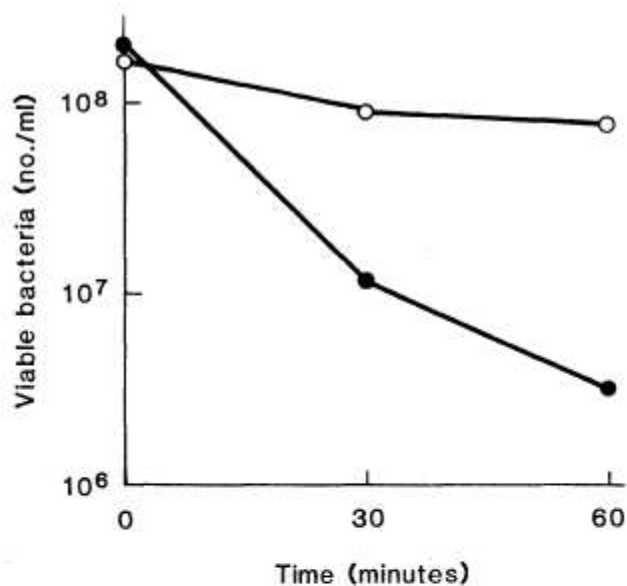


Fig. 2. Neutrophil killing defect induced by *Bordetella* extract. Human neutrophils ( $2 \times 10^7$  per milliliter) suspended in Hanks balanced salt solution were incubated for 5 minutes at  $37^\circ\text{C}$  with an equal volume of dialyzed *Bordetella* extract or dialysate control. The killing of *Staphylococcus aureus* 502A was assessed as described (12) by admixing  $5 \times 10^6$  neutrophils,  $2 \times 10^8$  bacteria, and 0.1 ml of pooled human serum in a total volume of 1 ml. Numbers of viable bacteria remaining were determined by plating dilutions of the incubation suspension removed at 0, 30, and 60 minutes. Each point represents the mean of quadruplicate determinations. Control tubes containing no neutrophils showed no change in bacterial count. Symbols: ○, *Bordetella*-treated neutrophils; ●, control neutrophils.

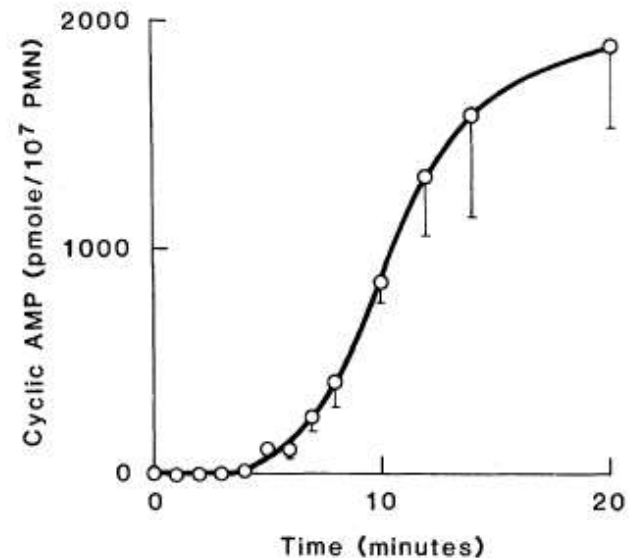


Fig. 3. Accumulation of cyclic AMP in human neutrophils (PMN) incubated with dialyzed *Bordetella* extract. Neutrophils,  $10^7$  per milliliter in Hanks balanced salt solution, were incubated at  $37^\circ\text{C}$  with equal volumes of dialyzed *Bordetella* extract (protein content,  $520 \mu\text{g/ml}$ ) for the times shown. Total cyclic AMP was determined as described (16). Values shown represent the means and standard error of seven separate (duplicate) determinations with neutrophils from four different donors. Normal neutrophils contain 2 to 5 pmole of cyclic AMP per  $10^7$  cells, and these amounts do not change during control incubations. Separate experiments (not shown), in which neutrophil pellets were obtained by brief centrifugation after incubation, indicated that  $> 90$  percent of the total recoverable cyclic AMP is associated with the cell pellet.

Nobody doubts that antibodies against PT save infant`s lives

= PT needs to be in the aP vaccine

but:

*B. parapertussis* does not need PT to  
cause whooping cough

and

Adenylate cyclase toxin is critical for  
colonization *by Bordetella pertussis*

Goodwin MS, Weiss AA. (1990) *Infect Immun.* 58:3445-7

Khelef N, Sakamoto H, Guiso N. (1992) *Microb. Pathog.* 12:227-35

# The bug really needs ACT for knocking down innate immunity:

- *All Bordetellae* pathogenic to mammals produce adenylate cyclase toxin-hemolysin (Except for certain *B. bronchispetica* lineages)
- ACT sequences are **highly conserved in *B. pertussis* isolates**
- **Strains not producing ACT have not been isolated** from patients, so far (in contrast to PT, FHA or pertactin, which all are dispensable)
- ACT is an extremely potent toxin that **knocks-down phagocytes in 30-60 seconds** (PT needs 30 min to get internalized into cells and effects manifest in 8- 12 h)
- ACT **instantaneously blocks oxidative burst of neutrophils** at pM conc. in 30 seconds
- ACT **blocks uptake of complement-opsonized particles** at pM conc.

# Not suprisingly, hence, ACT is a protective antigen

INFECTION AND IMMUNITY, Sept. 1993, p. 3583-3589  
0019-9567/93/091583-07\$02.00/0  
Copyright © 1993, American Society for Microbiology

Vol. 61, No. 9

INFECTION AND IMMUNITY, Sept. 1995, p. 3309-3315  
0019-9567/95/03091583-07\$02.00/0  
Copyright © 1995, American Society for Microbiology

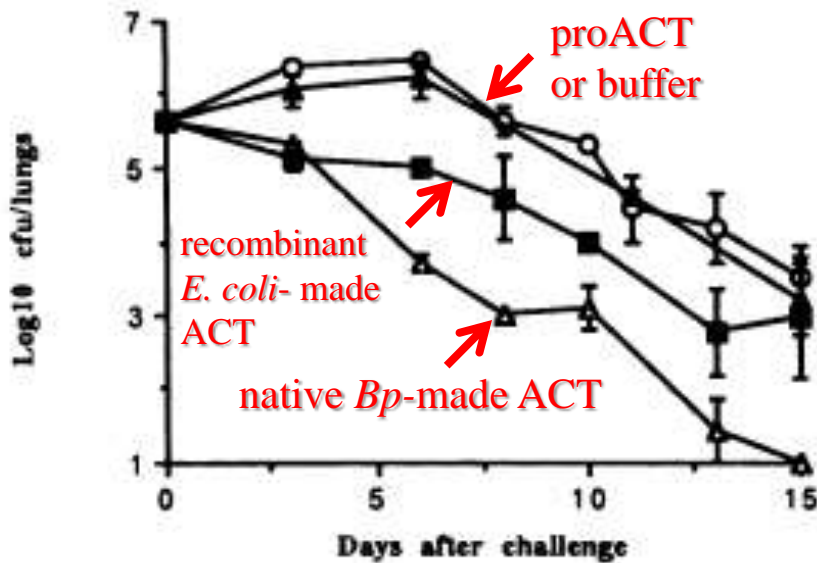
Vol. 63, No. 9

## CyaC-Mediated Activation Is Important Not Only for Toxic but Also for Protective Activities of *Bordetella pertussis* Adenylate Cyclase-Hemolysin

FOTINI BETSOU,<sup>1</sup> PETER ŠEBO,<sup>2</sup> AND NICOLE GUISO<sup>1\*</sup>

Unité de Bactériologie Moléculaire et Médicale<sup>1</sup> and Unité de Biochimie des Régulations Cellulaires,<sup>2</sup> Institut Pasteur, 28 rue du Dr. Roux, 75724 Paris Cedex 15, France

Received 8 April 1993/Returned for modification 7 May 1993/Accepted 1 June 1993



(at that time ACT samples contained LPS)

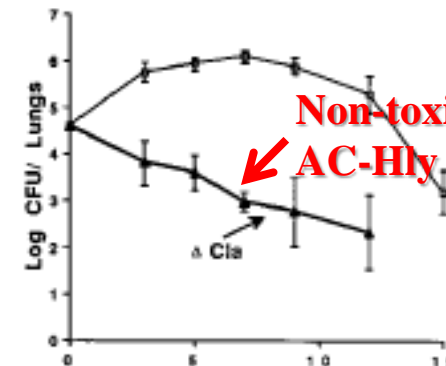
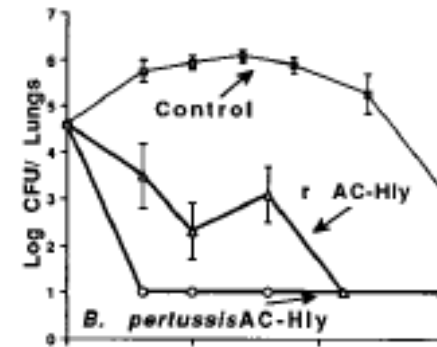
## The C-Terminal Domain Is Essential for Protective Activity of the *Bordetella pertussis* Adenylate Cyclase-Hemolysin

FOTINI BETSOU,<sup>1</sup> PETER ŠEBO,<sup>2†</sup> AND NICOLE GUISO<sup>1\*</sup>

Unité de Bactériologie Moléculaire et Médicale<sup>1</sup> and Unité de Biochimie des Régulations Cellulaires,<sup>2</sup> Institut Pasteur, 75724 Paris Cedex 15, France

Received

1995



# Highly purified CyaA-AC<sup>-</sup> protects on its own



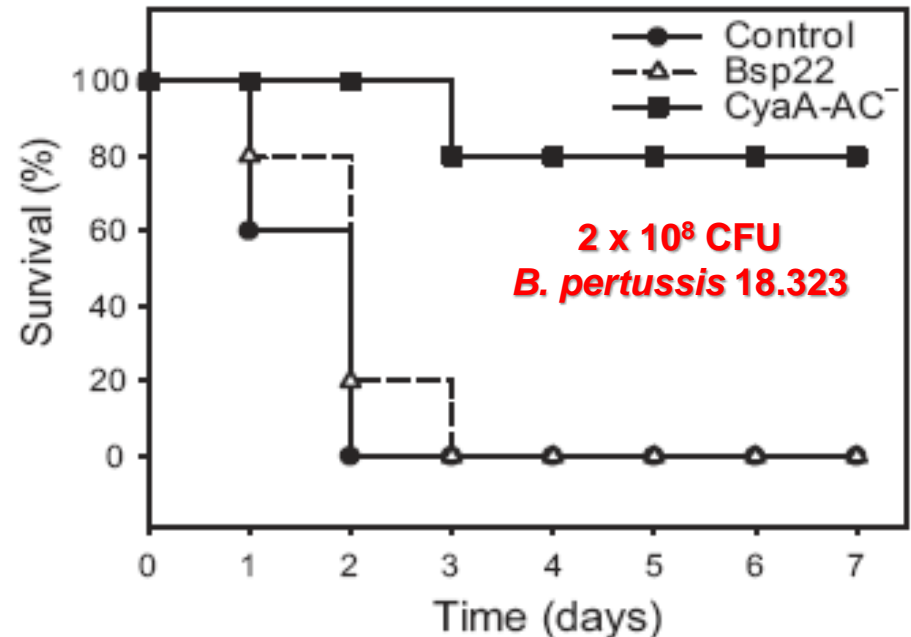
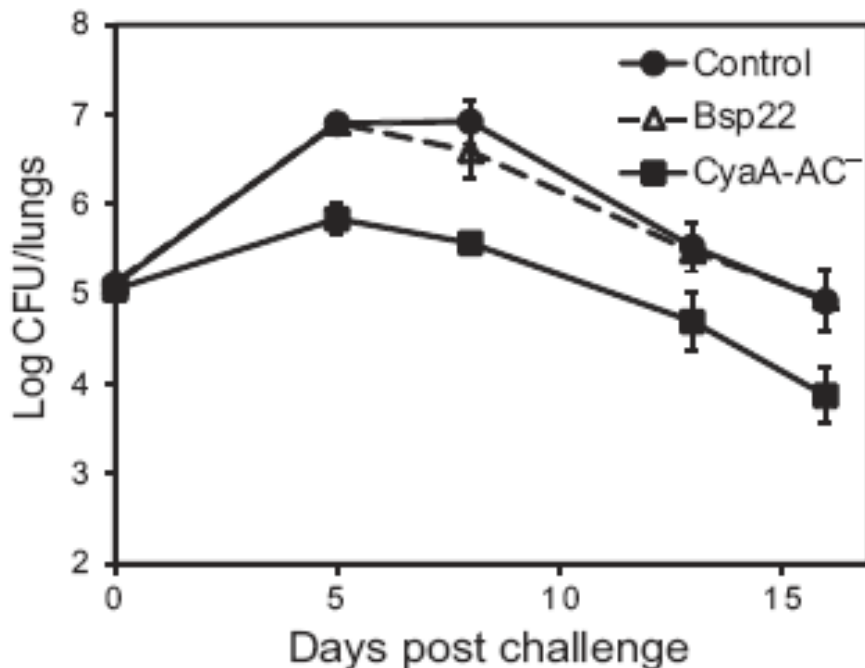
*Infection and Immunity* **81**:  
2761–2767 (2013)

The *Bordetella pertussis* Type III Secretion System Tip Complex Protein Bsp22 Is Not a Protective Antigen and Fails To Elicit Serum Antibody Responses during Infection of Humans and Mice

Rodrigo Villarino Romero,<sup>1</sup> Ilona Bibova,<sup>2</sup> Ondrej Cerny,<sup>2</sup> Branislav Vecerek,<sup>2</sup> Tomas Wald,<sup>2</sup> Oldrich Benada,<sup>2</sup> Jana Zavadilova,<sup>2</sup> Radim Osicka,<sup>2</sup> Peter Sebo<sup>2</sup>  
Institute of Microbiology of the ASCR, Prague, Czech Republic<sup>1</sup>; National Institute of Public Health, Prague, Czech Republic<sup>2</sup>



Poster: Villarino, Bibova *et al.*



# Evidence from other labs

- Hormozi K, Parton R, Coote J. **Adjuvant and protective properties of native and recombinant Bordetella pertussis adenylate cyclase toxin** preparations in mice. FEMS Immunol Med Microbiol 1999;23(4): 273-82
- Macdonald-Fyall J, Xing D, Corbel M, et al. **Adjuvanticity of native and detoxified adenylate cyclase toxin** of Bordetella pertussis towards co-administered antigens. Vaccine 2004;22(31-32):4270-81
- Orr B, Douce G, Baillie S, et al. **Adjuvant effects of adenylate cyclase toxin** of Bordetella pertussis after intranasal immunisation of mice. Vaccine 2007;25(1): 64-71
- Cheung GY, Xing D, Prior S, et al. **Effect of different forms of adenylate cyclase toxin of Bordetella pertussis on protection afforded by an acellular pertussis vaccine in a murine model.** Infect Immun 2006;74(12):6797-805

# Addition of CyaA-AC<sup>-</sup> improves performance of the aP vaccine

INFECTION AND IMMUNITY, Dec. 2006, p. 6797-6805  
0019-9567/06/\$08.00+0 doi:10.1128/IAI.01104-06  
Copyright © 2006, American Society for Microbiology. All Rights Reserved.

Vol. 74, No. 12

**one-eighth of human dose of DTaP/ACV**  
(Infanrix, GSK) + **CyaA-AC<sup>-</sup>** 2 x i.p.  
challenged with  $4 \times 10^6$  *B. pertussis* 18.323 i.n.

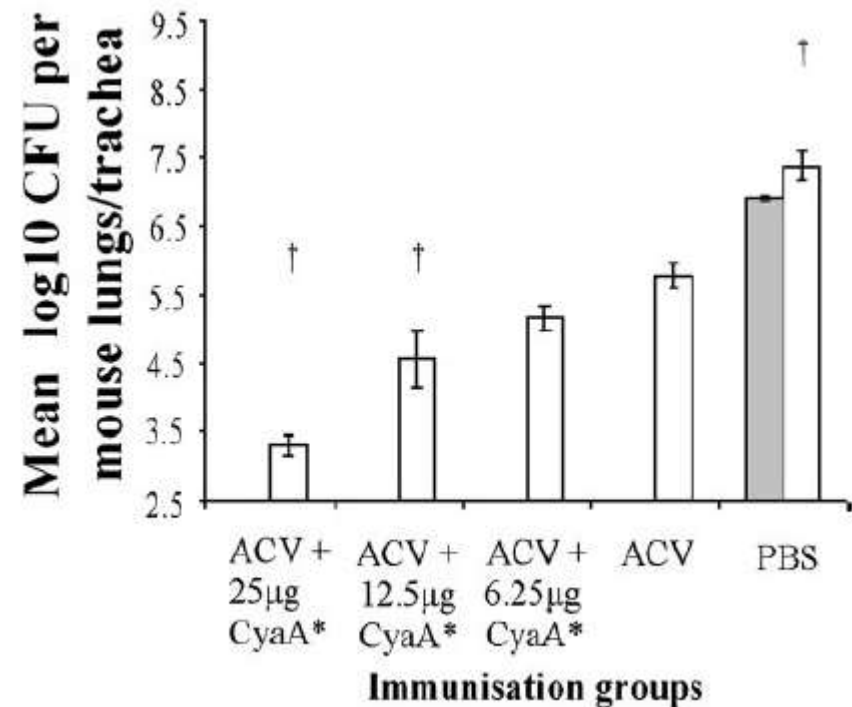
## Effect of Different Forms of Adenylate Cyclase Toxin of *Bordetella pertussis* on Protection Afforded by an Acellular Pertussis Vaccine in a Murine Model<sup>V</sup>

Gordon Y. C. Cheung,<sup>1</sup> Dorothy Xing,<sup>2</sup> Sandra Prior,<sup>2</sup> Michael J. Corbel,<sup>2</sup>  
Roger Parton,<sup>1</sup> and John G. Coote<sup>1\*</sup>

Division of Infection and Immunity, Glasgow Biomedical Research Centre, University of Glasgow, Glasgow,<sup>1</sup> and Division of Bacteriology, National Institute of Biological Standards and Control, South Mimms, Hertfordshire,<sup>2</sup> United Kingdom

Received 14 July 2006/Accepted 12 September 2006

Four recombinant forms of the cell-invasive adenylate cyclase toxin (CyaA) of *Bordetella pertussis* were compared for the ability to enhance protection against *B. pertussis* in mice when coadministered with an acellular pertussis vaccine (ACV). The four forms were as follows: fully functional CyaA, a CyaA form lacking adenylate cyclase enzymatic activity (CyaA\*), and the nonacylated forms of these toxins, i.e., proCyaA and proCyaA\*, respectively. None of these forms alone conferred significant ( $P > 0.05$ ) protection against *B. pertussis* in a murine intranasal challenge model. Mice immunized with ACV alone showed significant ( $P < 0.05$ ) reductions in bacterial numbers in the lungs after intranasal challenge compared with those for control mice. When administered with ACV, both CyaA and CyaA\* further reduced bacterial numbers in the lungs of mice after intranasal challenge compared with those for ACV-immunized mice, but the enhanced protection was only significant ( $P < 0.05$ ) with CyaA\*. Coadministration of CyaA\* with ACV caused a significant ( $P < 0.05$ ) increase in immunoglobulin G2a antibody levels against pertactin compared with those in mice immunized with ACV alone. Spleen cells from mice immunized with ACV plus CyaA\* secreted larger amounts of interleukin-5 (IL-5), IL-6, gamma interferon (IFN- $\gamma$ ), and granulocyte-macrophage colony-stimulating factor (GM-CSF) than did cells from mice immunized with ACV plus CyaA or ACV alone after stimulation in vitro with a mixture of *B. pertussis* antigens. Spleen cells from mice immunized with ACV plus CyaA\* also secreted larger amounts of IFN- $\gamma$  and GM-CSF than did cells from mice immunized with CyaA\* alone after stimulation in vitro with CyaA\*. Macrophages from mice immunized with ACV plus CyaA\* produced significantly ( $P < 0.05$ ) higher levels of nitric oxide than did macrophages from mice immunized with CyaA\* alone, ACV alone, or ACV plus CyaA after stimulation in vitro with a mixture of *B. pertussis* antigens or heat-killed *B. pertussis* cells. These data suggest that the enhancement of protection provided by CyaA\* was due to an augmentation of both Th1 and Th2 immune responses to *B. pertussis* antigens.



# ACT relevance for pertussis vaccines

(Sebo P. *et al.* Expert Rev. Vaccines 13(10), 1215–1227 (2014))

- **ACT is a protective antigen** in the i.n. or aerosol challenge model
- **anti-ACT antibodies are common and abundant in convalescent patients**
- documented adjuvant effect of the nonenzymatic AC<sup>-</sup> toxoid on co-administered B. pertussis antigens
- **ACT polarizes T cell responses towards Th1** even when administered with alum!
- **AC<sup>-</sup> toxoid drives maturation of DC and** when mixed with aP vaccine antigens and alum AC toxoid **could partly shift the polarization of the immune response from a typical Th2 type, seen with the aP vaccine in mice, to a mixed Th1/Th2 type of response....**
  - Cheung GY, Xing D, Prior S, et al. Effect of different forms of adenylate cyclase toxin of Bordetella pertussis on protection afforded by an acellular pertussis vaccine in a murine model. Infect Immun 2006;74(12):6797-805



# We shall know in a few years...



PRESS RELEASE

Paris and Toulouse, February 2<sup>nd</sup>, 2015

## GENTICEL LICENSES VAXICLASE PLATFORM TO SERUM INSTITUTE OF INDIA FOR USE IN PERTUSSIS VACCINES

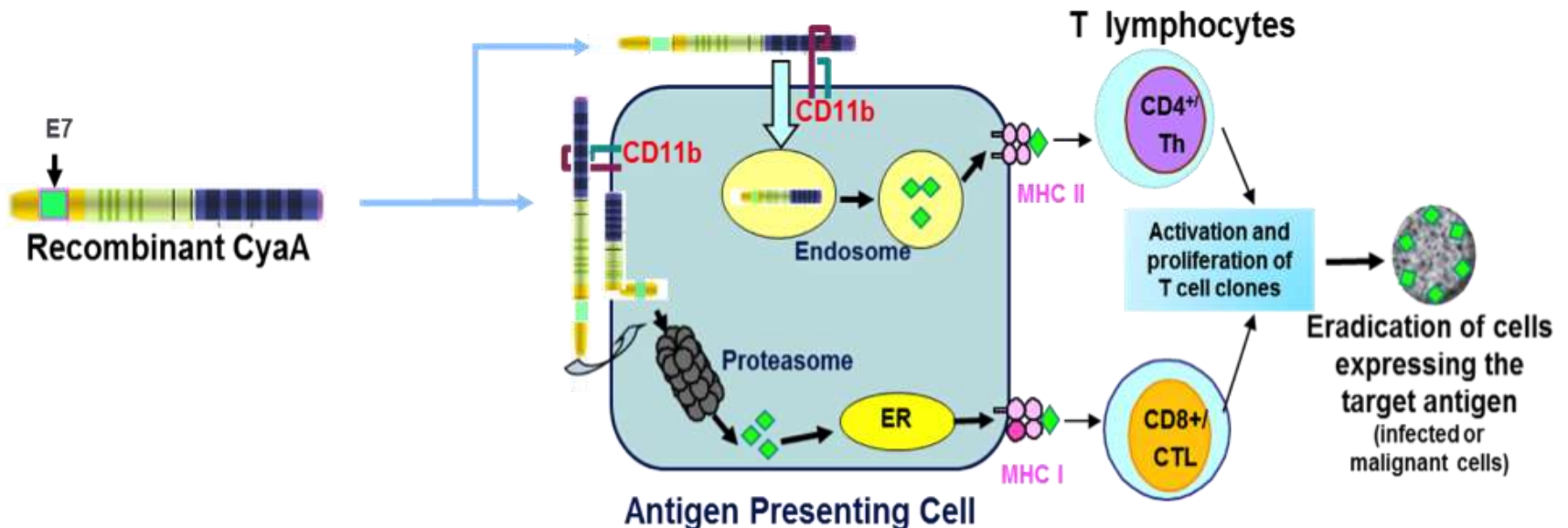
- Serum Institute of India, world's largest producer of vaccines, to evaluate Vaxiclase for use in multivalent vaccines containing pertussis antigens
- Preclinical stage license agreement entitles Genticel to up to \$57 million in upfront & milestones payments plus single digit royalties on net sales

GENTICEL (Euronext Paris and Brussels: FR0011790542 - GTCL), a French biotechnology company and leading developer of therapeutic vaccines, today announces that it has licensed its Vaxiclase technology to Serum Institute of India Ltd. (SIIL), for use as a component in acellular multivalent combination vaccines including pertussis antigens.

The license granted by Genticel to SIIL provides the Vaxiclase platform for inclusion in multivalent vaccines that also protect against *Bordetella pertussis*, the causative agent of whooping cough. The license covers all countries of the world except major pharmaceutical markets, in particular the USA, Canada, New Zealand, Australia, Japan, Israel, Turkey and greater Europe.

**Vaccines that are built with the CyaA vector are chimeric recombinant proteins consisting of the CyaA protein and the antigen of choice.**

- 1: Sebo P, Fayolle C, d'Andria O, Ladant D, Leclerc C, Ullmann A. (1995) *Infect Immun.* 63(10):3851-7.
- 2: Fayolle C, Sebo P, Ladant D, Ullmann A, Leclerc C. (1996) *J Immunol.* 156(12):4697-706.
3. Saron MF, Fayolle C, Sebo P, Ladant D, Ullmann A, Leclerc C. *Proc Natl Acad Sci U S A.* 1997;94(7):3314-9.





**GENTICEL**  
GENTLE T CELL VACCINES

[www.genticel.com](http://www.genticel.com)

**September 2012 - Genticel S.A. completed Phase I clinical trial for HPV16/18-induced cervical carcinoma**

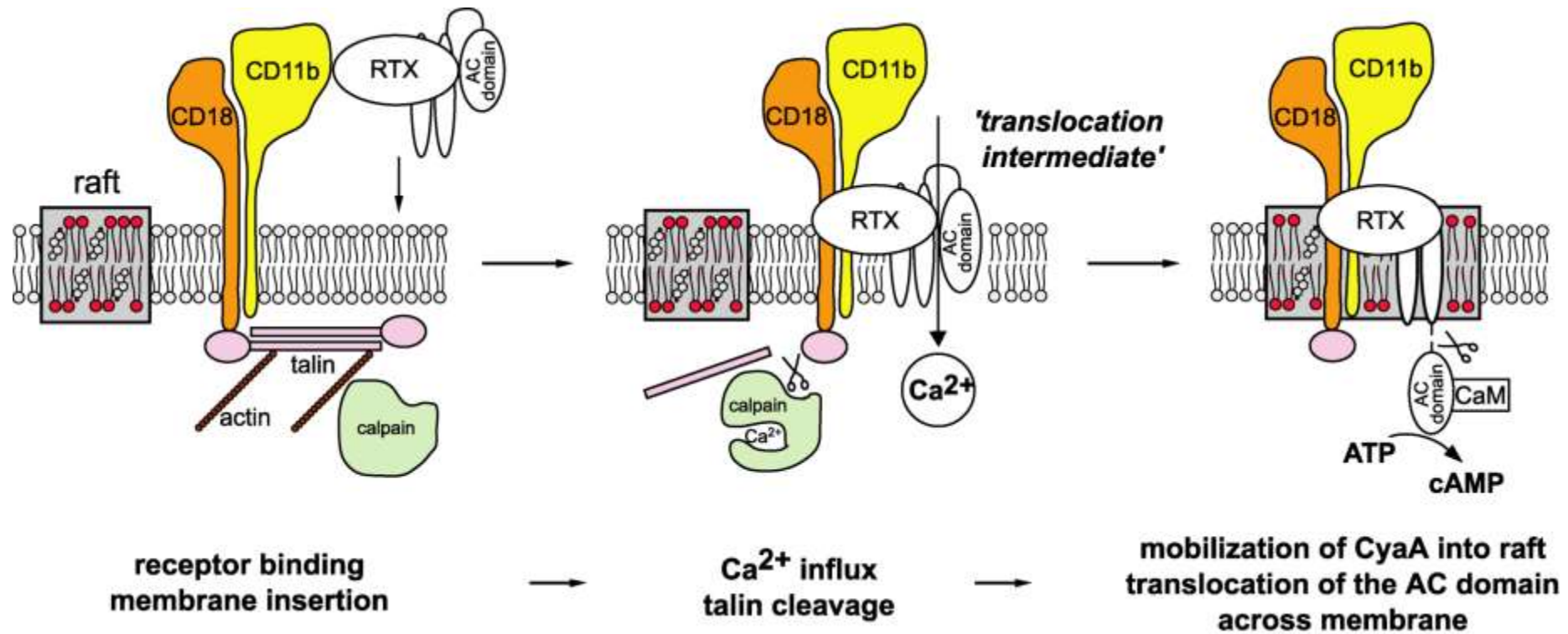
**Using a cGMP batch of the adenylate cyclase (CyaA-AC<sup>-</sup>) toxoid for delivery of HPV E7 antigen as immunotherapeutic vaccine**

**safe, immunogenic, inducing CD8<sup>+</sup> CTLs and HPV 16/18 virus load reduction demonstrated**

**Entered phase II trial = will be of interest to see pertussis incidence in CyaA-E7 toxoid treated woman...**

**IPO on April 4, 2014 at Euronext Paris and Brussels - 34 millions Euro**

# Adenylate cyclase toxin hijacks the $\beta_2$ integrin receptor into lipid rafts to accomplish membrane translocation in two steps



Bumba et al. (2010). PLoS Pathog 6(5): e1000901.

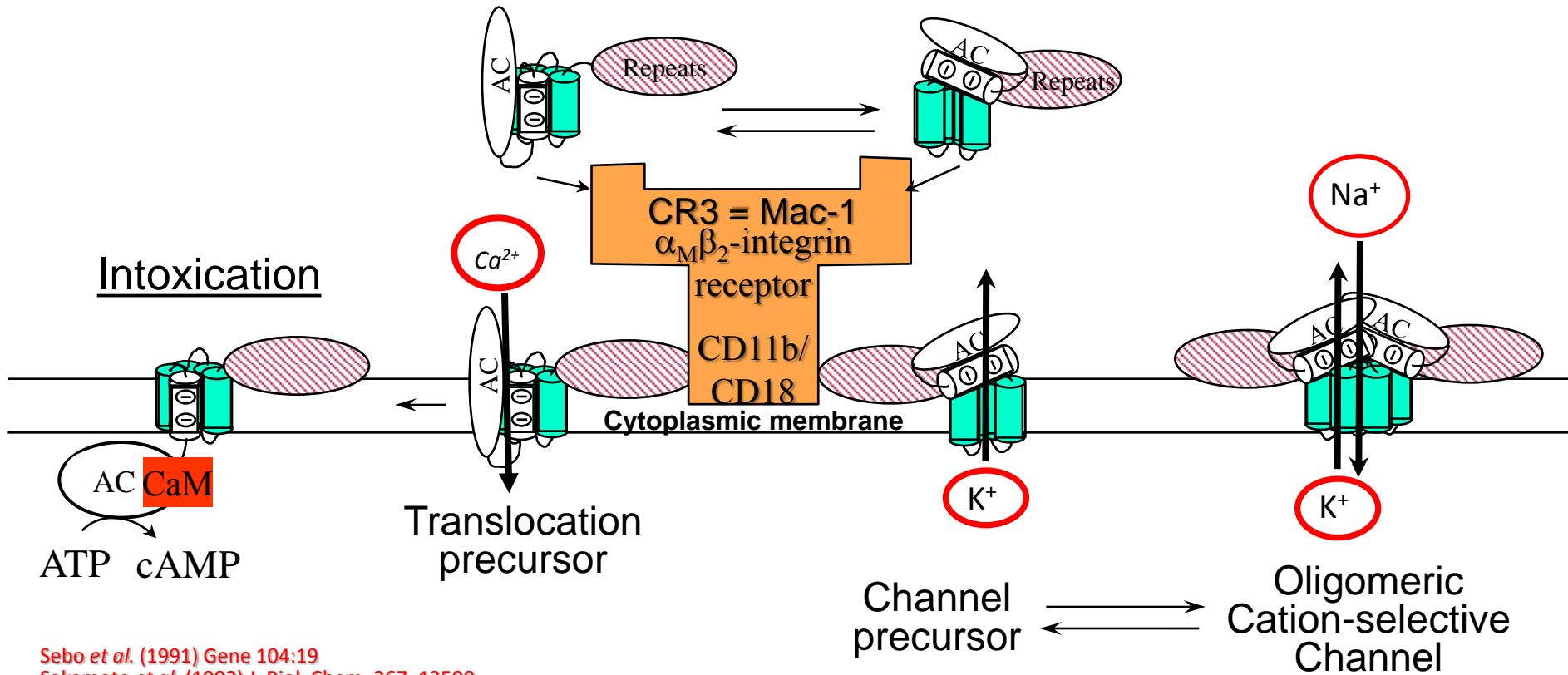
**CyaA-AC<sup>-</sup> higher concentrations  
induces maturation of DC through LPS-independent  
TLR4 and TRIF signaling..**

## **Antigen Targeting to CD11b<sup>+</sup> Dendritic Cells in Association with TLR4/TRIF Signaling Promotes Strong CD8<sup>+</sup> T Cell Responses**

- CyaA binds CD11b<sup>+</sup> dendritic cells (DCs) and induces their maturation...
- DCs sense CyaA through the TLR4/Toll/IL-1R domain–containing adapter-inducing IFN- $\beta$  pathway, independent of the presence of LPS
- leading to the induction of strong immune responses.

**Dadaglio *et al.* (2014) J. Immunol. 193: 1787–1798.**

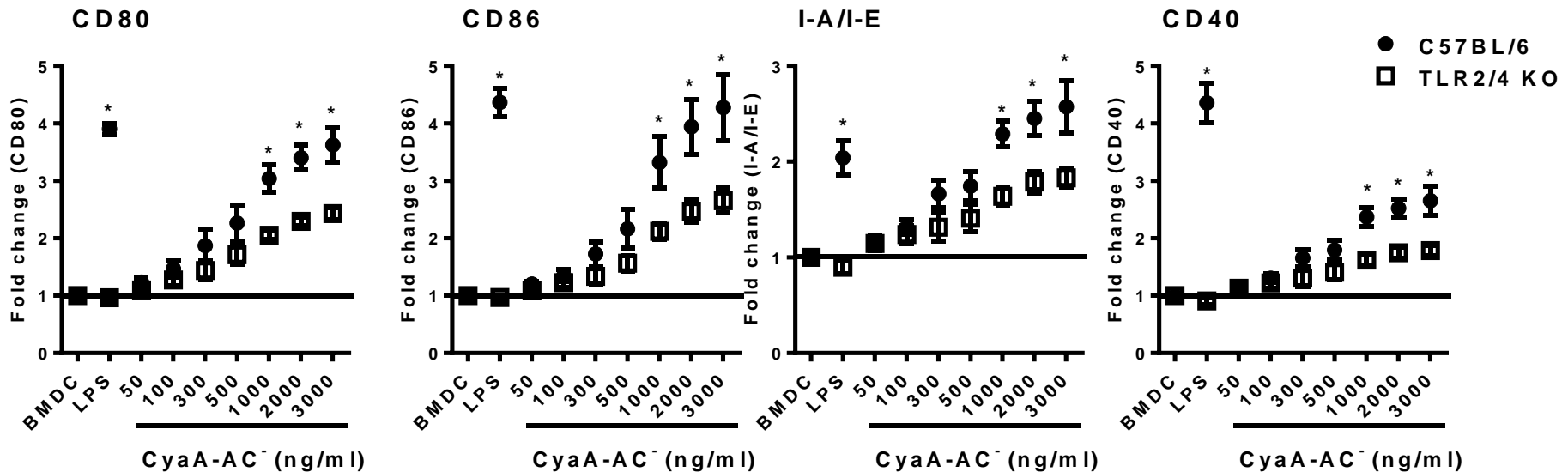
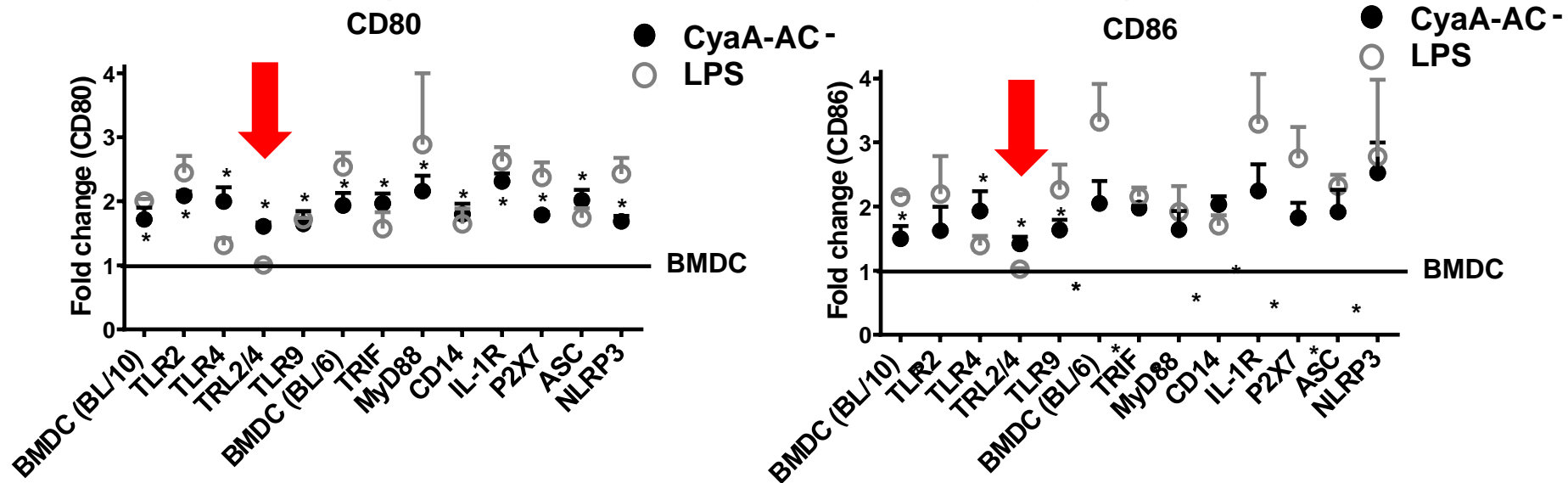
# The three cytotoxic activities of ACT adenylate cyclase toxin & pore-forming hemolysin/Cytolysin



Sebo *et al.* (1991) *Gene* 104:19  
 Sakamoto *et al.* (1992) *J. Biol. Chem.* 267, 13598  
 Benz *et al.* (1994) *J. Biol. Chem.* 269, 27231  
 Hackett *et al.* (1995) *J. Biol. Chem.* 270, 20250  
 Gray *et al.* (1998) *J. Biol. Chem.* 273, 18260  
 Osickova *et al.* (1999) *J. Biol. Chem.* 274, 37644  
 Basler *et al.* (2007) *J. Biol. Chem.* 282, 12419  
 Fiser R. *et al.* (2007) *J. Biol. Chem.* 282, 2808  
 Osickova *et al.* (2010) *Mol. Microbiol.* 75:15450-1562

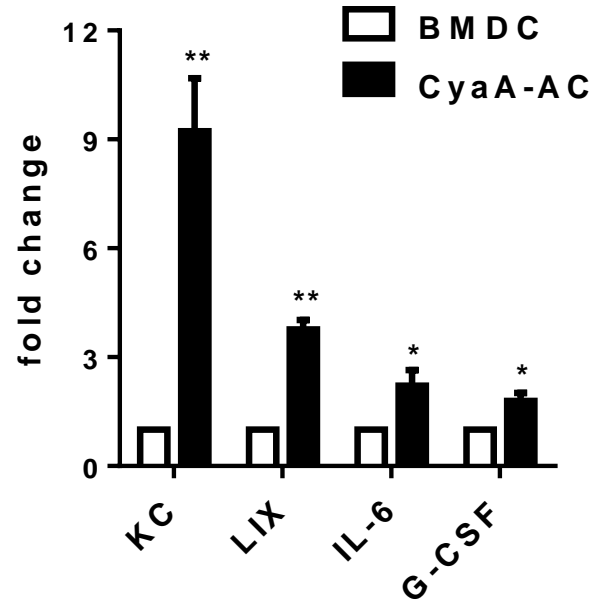
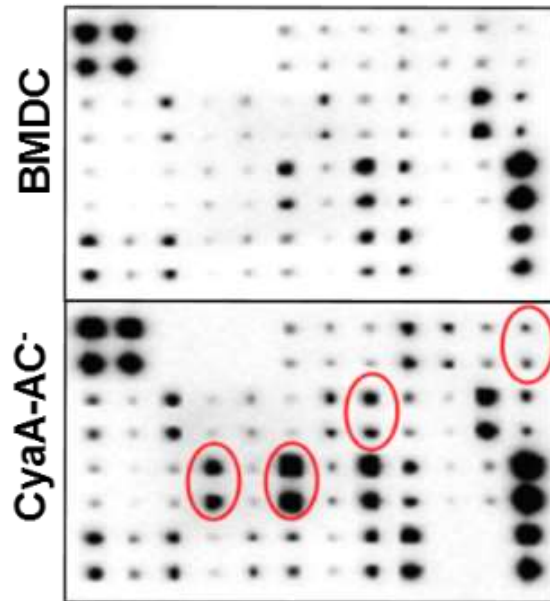
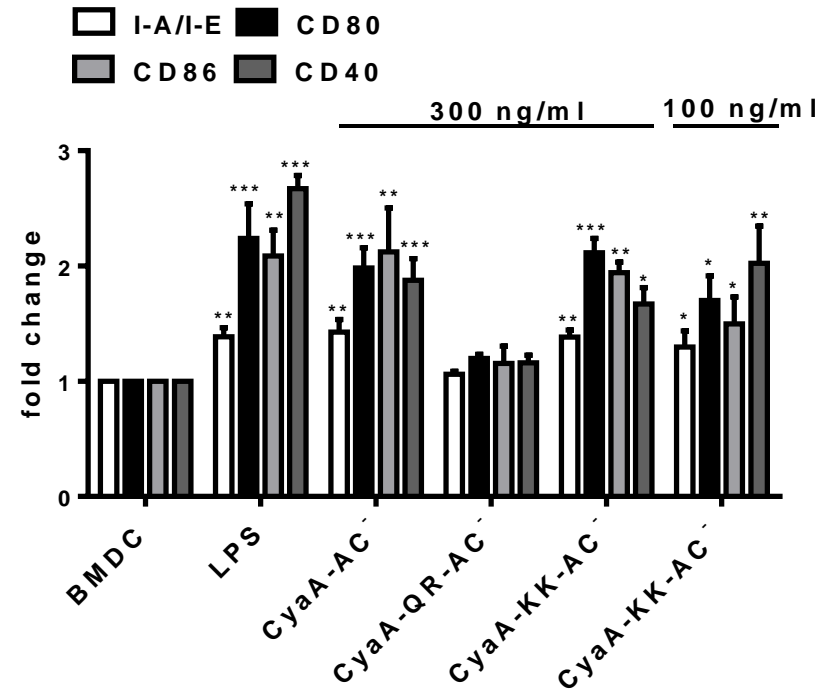
# However, already cell permeabilization by the Pore-forming activity by low amounts of CyaA-AC<sup>-</sup> induces maturation of DC

(Svedova et al. 2015 Immun Cell Biol, in press)



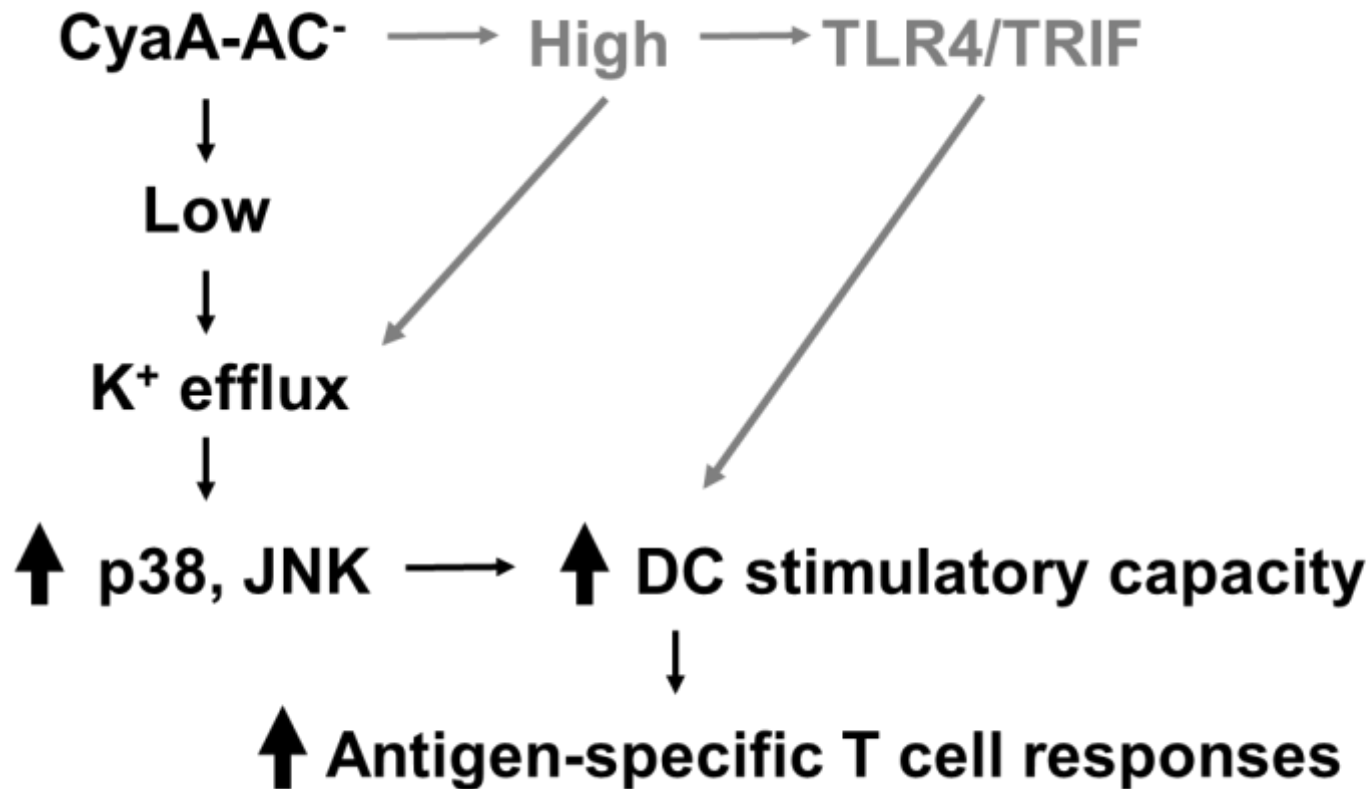
# Toxoid activates DC in function of Pore-forming activity (K<sup>+</sup> efflux)

CyaA mutants	Ca <sup>2+</sup> influx	Specific cell-permeabilizing activity (K <sup>+</sup> efflux)
CyaA-AC <sup>-</sup>	+++	++
CyaA-E570Q-K860R-AC	+++	-/+
CyaA-E509K-E516K-AC	-/+	++++





The CyaA-AC<sup>-</sup> Toxoid primes activation of DC by cell-permeabilization, causing K<sup>+</sup> efflux and p38 MAPK activation. At higher toxoid concentrations, CyaA-mediated clustering of CD11b/CD18 with TLR4 and TRIF signaling occurs

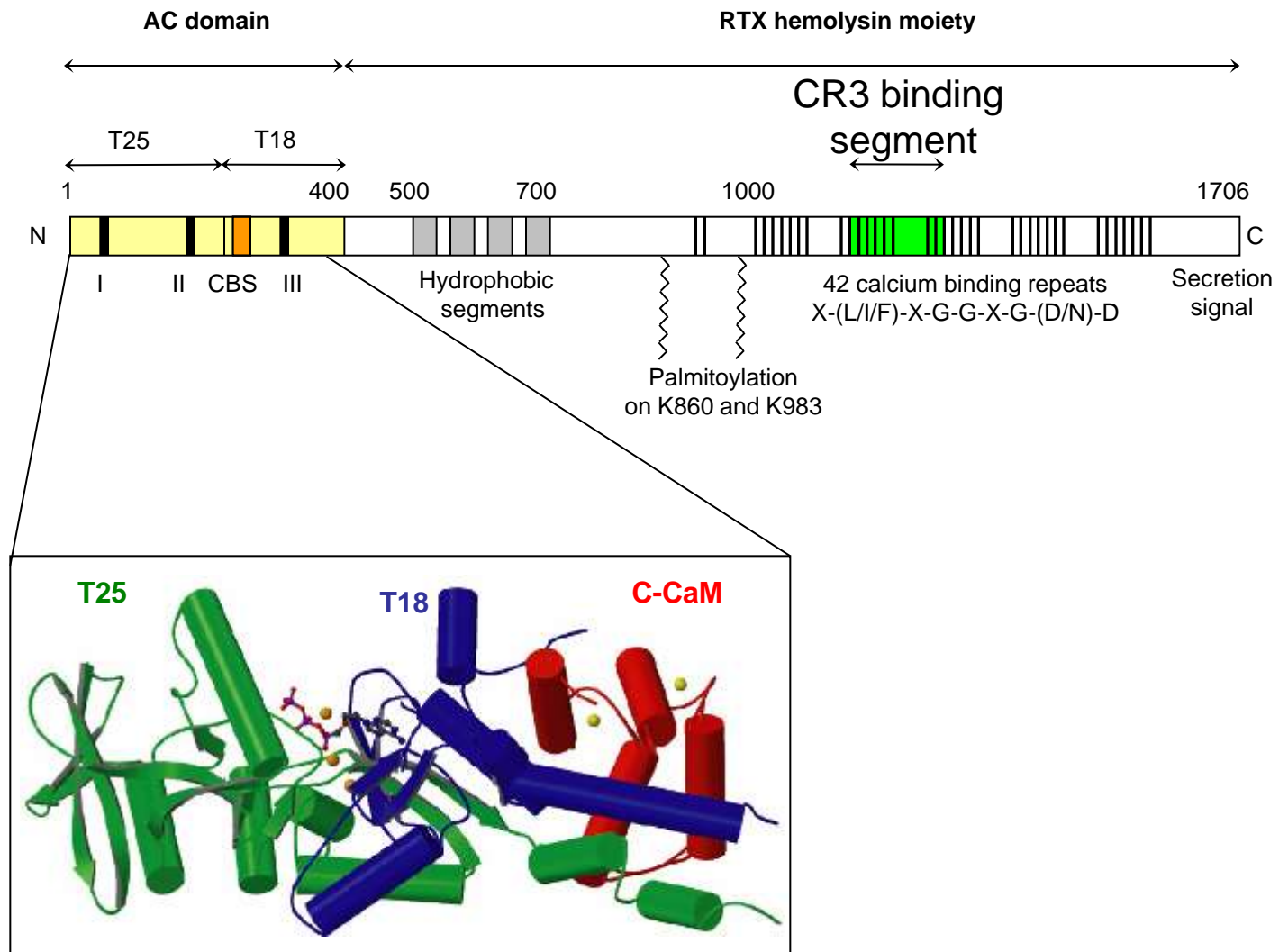


Svedova et al., (2015) Immunology and Cell Biology in press

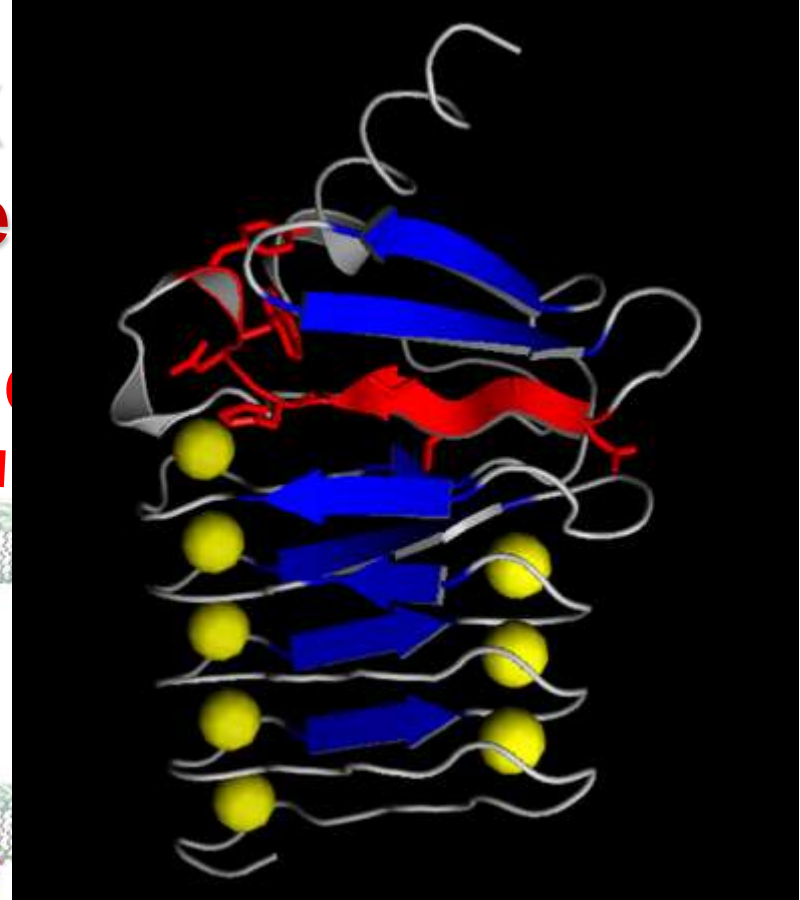
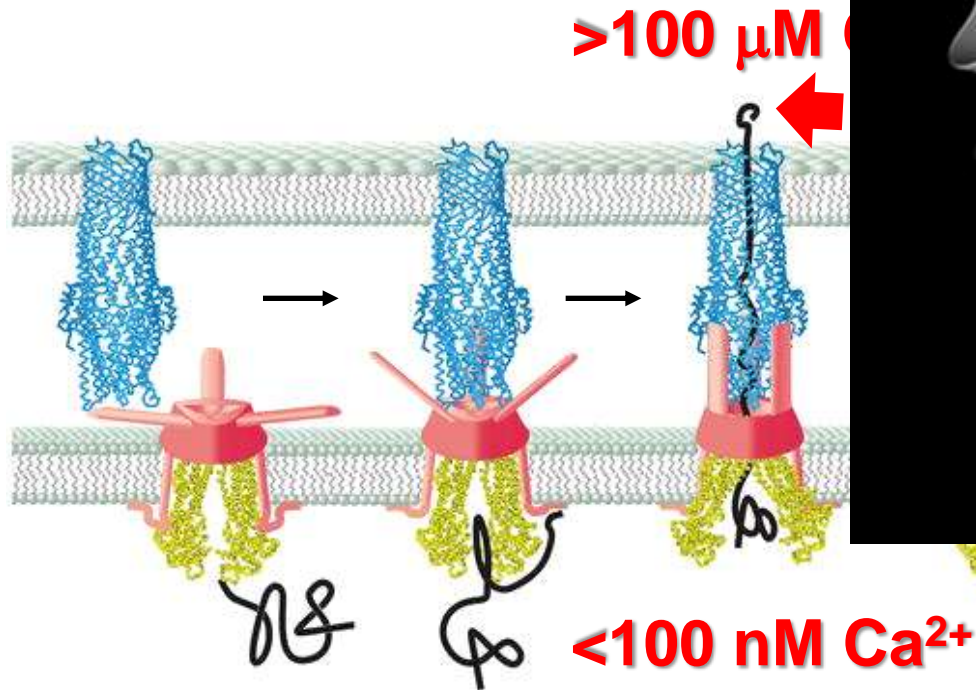
So, why is ACT so much relevant  
for pertussis vaccines?

What does it do and how does it  
work?

# Adenylate cyclase toxin - cytolysin



# ACT is an RTX secreted by a type

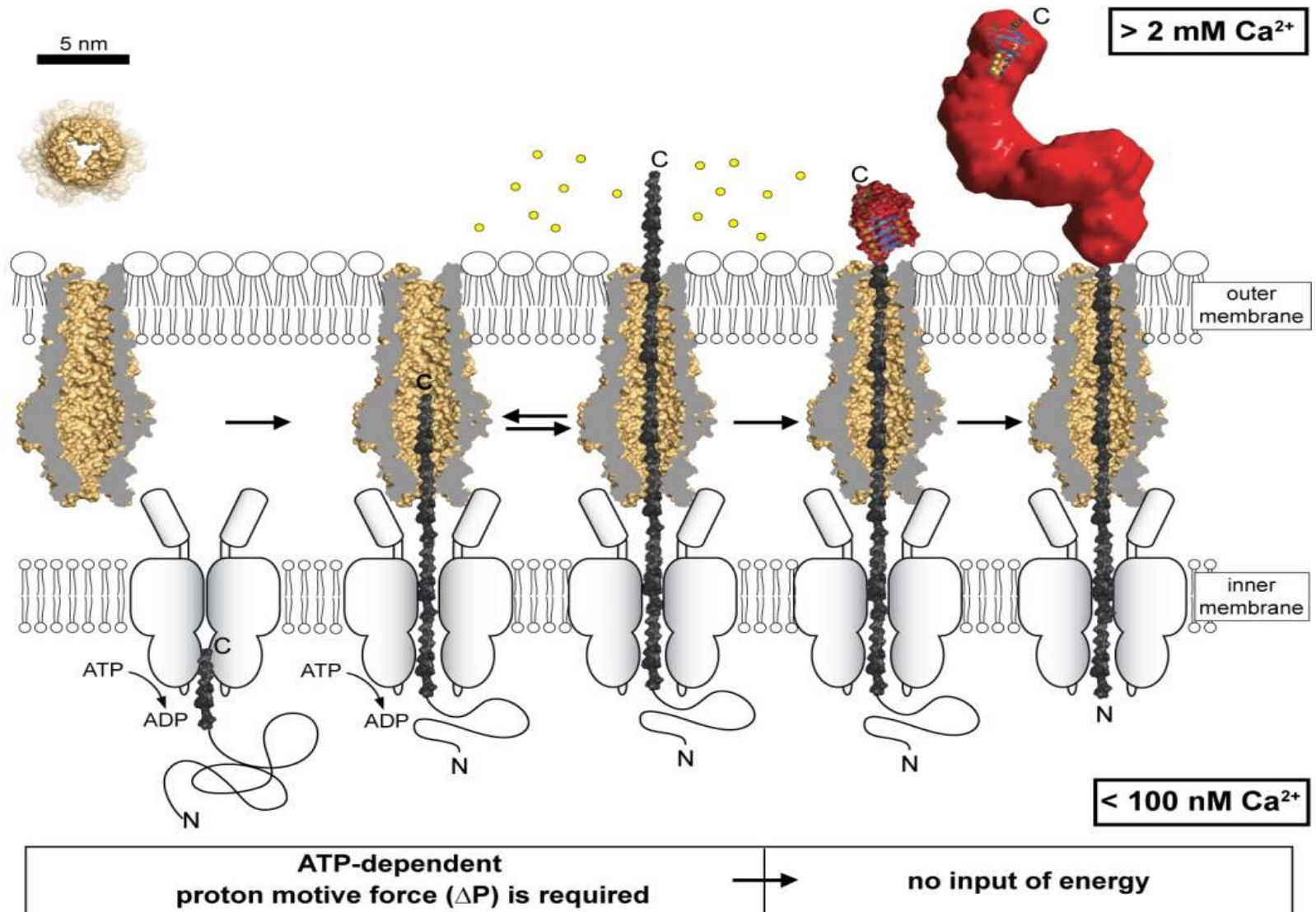


Need to unfold and refold  
on the way to target...



Lád'a

# Calcium-driven formation of an intramolecular Brownian ratchet directs movement of large RTX proteins through type I secretion system conduits



# ACT/cAMP signaling breaks the hell loose... and supresses TLR signaling of the bug...



## signal transduction events:

NF- $\kappa$ B,  $\downarrow$  MAPK – p38, ERK, JNK

## expression and

## upregulation of TLR:

TLR1-6, 9, TLR4, TLR2

mucin: MUC2, MUC5AC  $\uparrow$

## other soluble factors:

$\downarrow$  O<sub>2</sub><sup>-</sup>, NO,  $\uparrow$  PGE2

$\downarrow$  ciliary beating

## defensins and other antimicrobial peptides:

h $\beta$ defensin2,

$\downarrow$   $\beta$ defensin1,

$\downarrow$  cathelicidin

AEC



cAMP

other cells

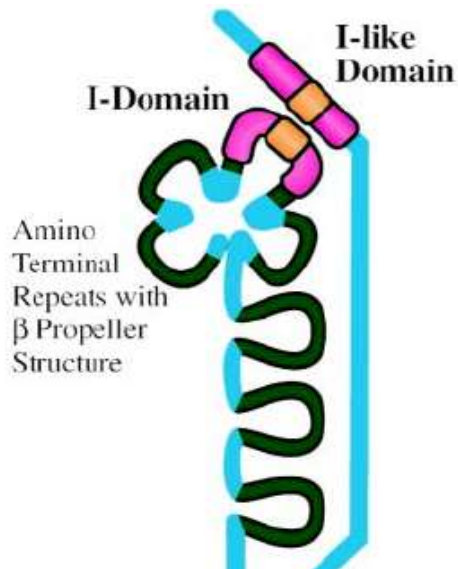
## cytokine and chemokines:

IL-1 $\alpha$ ,  $\uparrow$  IL-1 $\beta$ ,  $\uparrow$  IL-6,  $\uparrow$  IL-8,  
 $\uparrow$  IL-10,  $\downarrow$  TNF $\alpha$ ,  $\downarrow$  IFN $\beta$ , TGF- $\beta$ ,  
 $\downarrow$  GM-CSF, MCP-1,  $\downarrow$  MIP-1 $\alpha$ ,  
RANTES,..

## expression of costimulatory x

inhibitory molecules:  $\uparrow$  CD80, CD86,  $\downarrow$   
CD40,  $\downarrow$  CD54, B7-H2, B7-H3 x  $\uparrow$  FasL,  
PD-L1, PD-L2

# ACT targets myeloid phagocytes bearing $\alpha_M\beta_2$ integrin CD11b/CD18



- $\beta_2$  subfamily
- complement receptor 3 (CR3), Mac-1, Mo-1,  $\alpha_M\beta_2$
- monocytes, granulocytes, macrophages, NK cells, neutrophils and **dendritic cells**, certain B cell subtypes



**ACT first recognizes N-linked glycans of CD11b/CD18**

Morova et al. (2008) PNAS 105, 5355

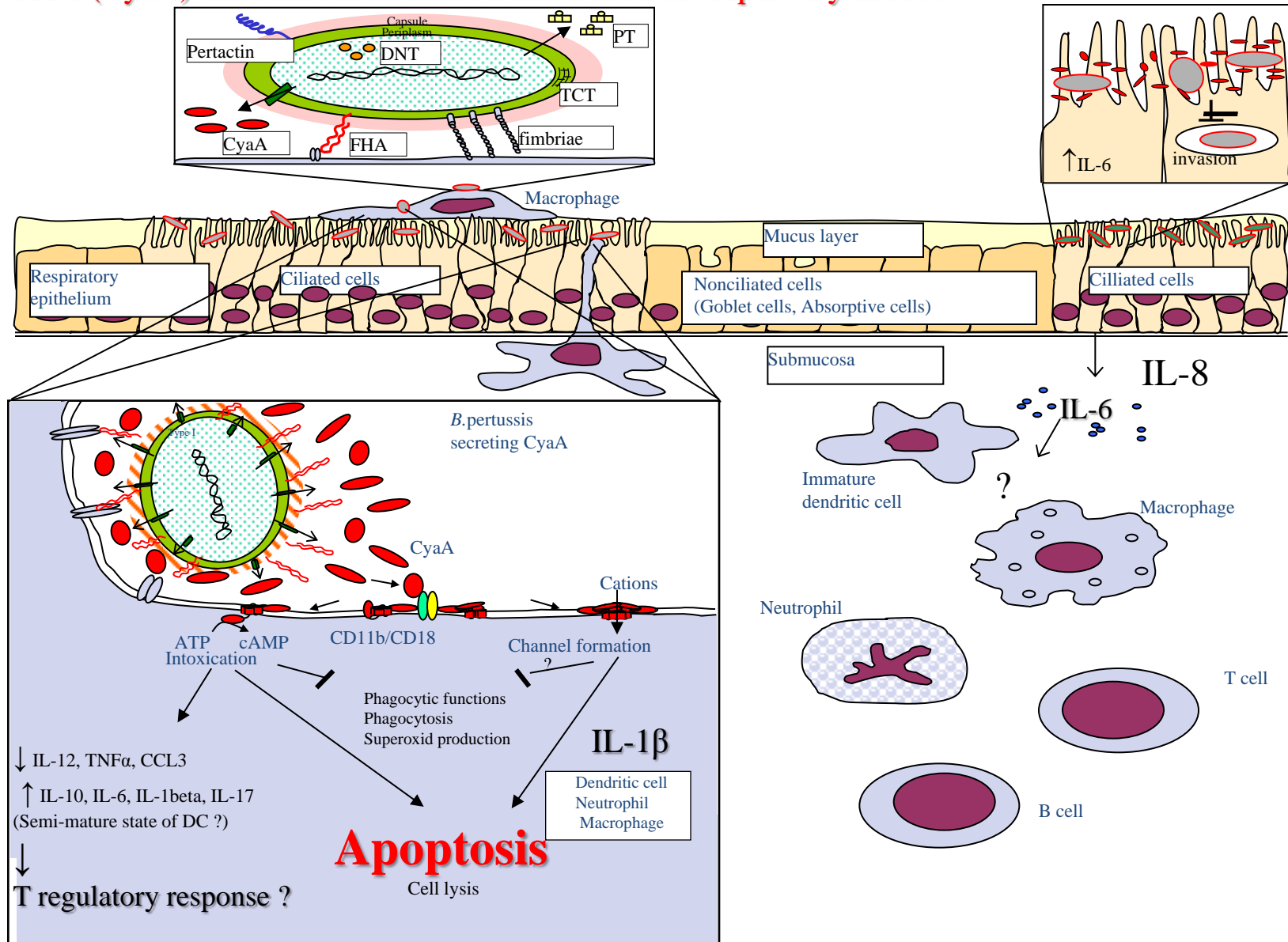
$\alpha$  subunit

$\beta$  subunit

Guermontprez et al. 2001, J Exp. Med.

# the Yin: ACT as a SWIFT SABOTEUR

low ACT (CyaA) concentrations make a difference on respiratory mucosa...



Osičková *et al.*, (1999) *J. Biol. Chem.* 274, 37644

Vojtová *et al.*, 2006, *Curr. Op.. Microbiol.* 9, 69-75

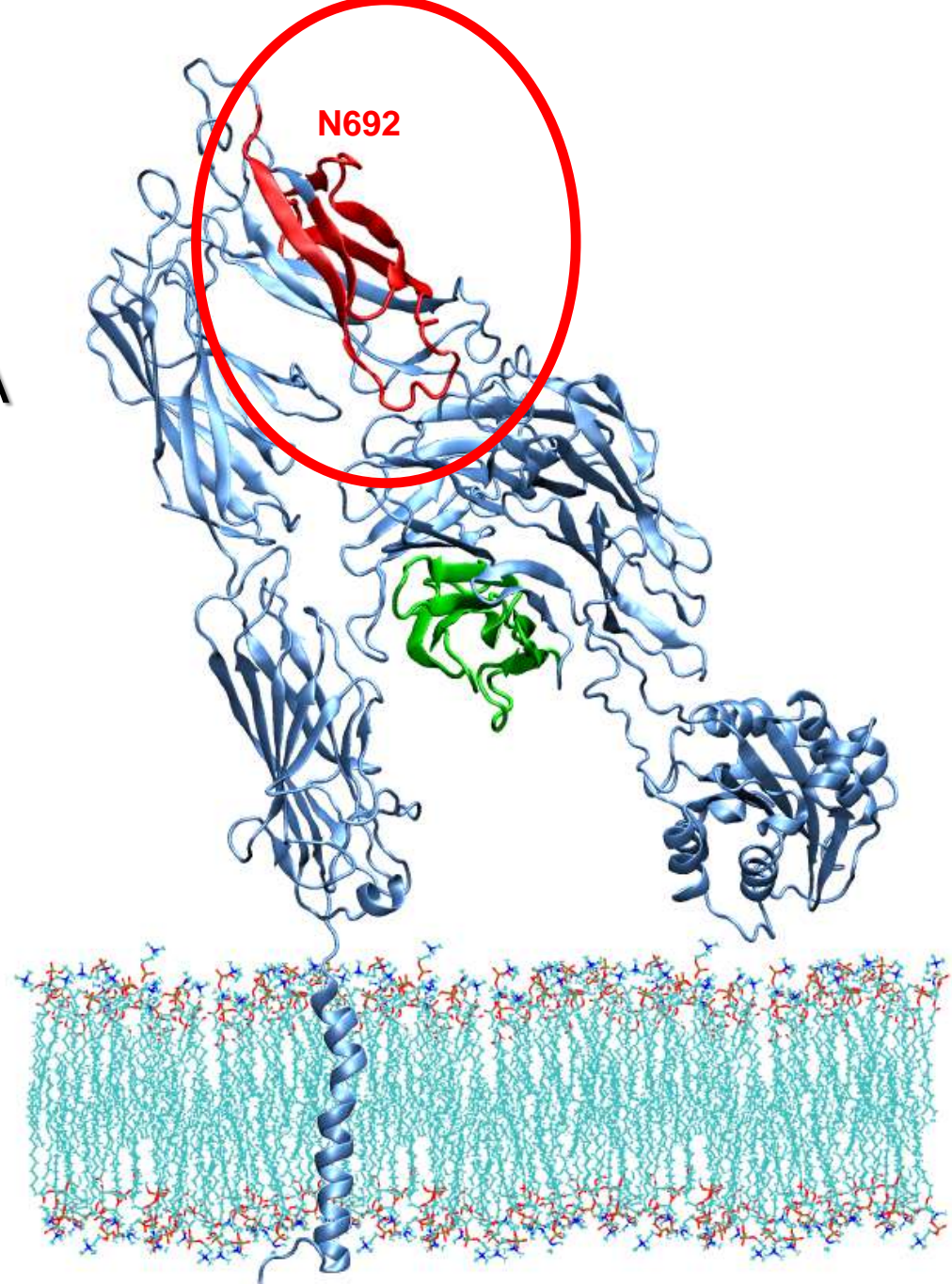


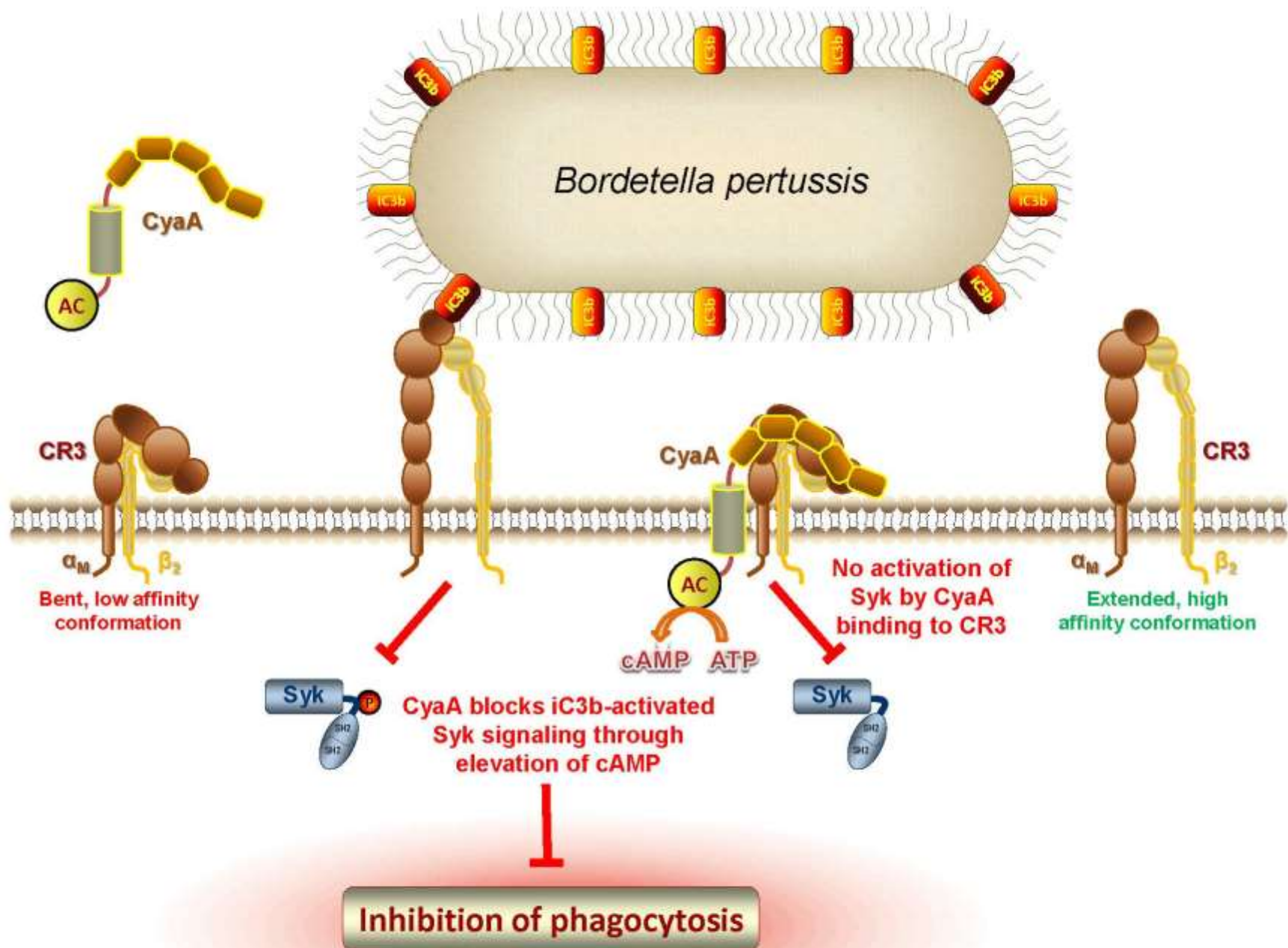
# Signaling disarming phagocytes

Proteinaceous segments  
Specifically involved in CyaA  
binding:

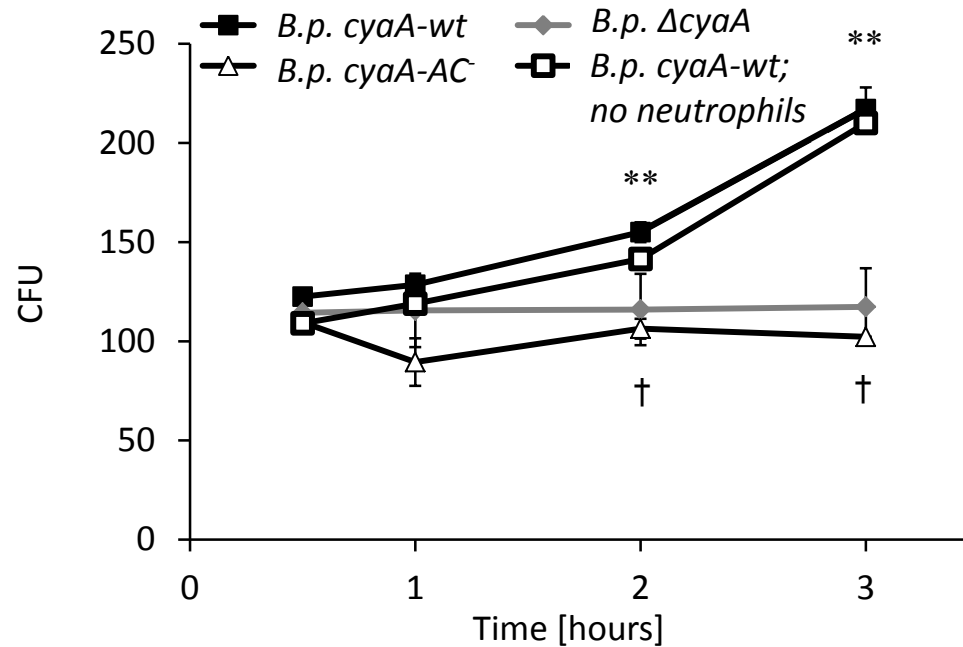
**CD11b – residues 614-682**

**CD11b - residues 342-424**



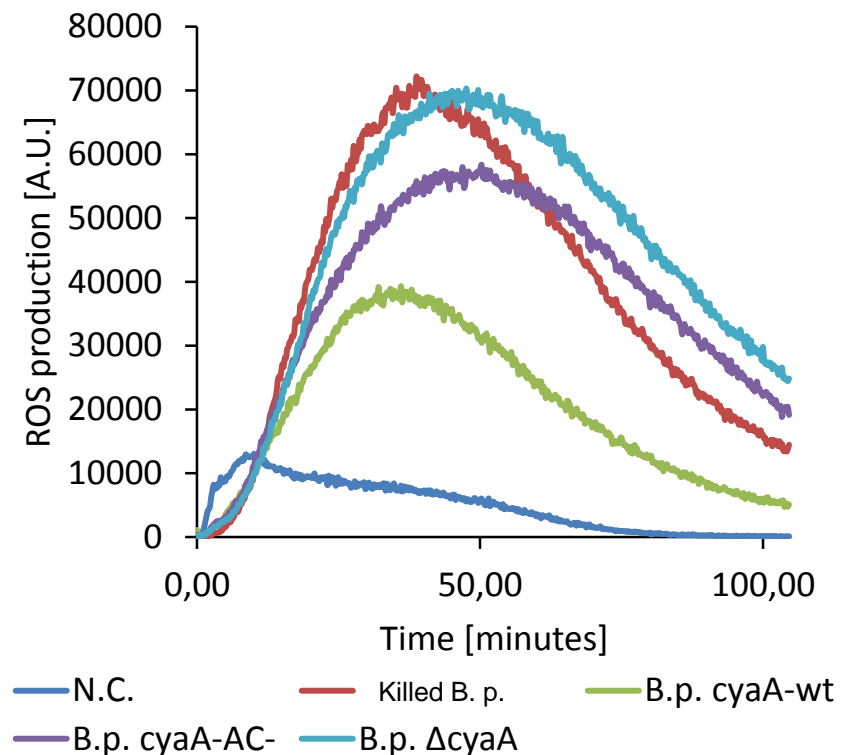
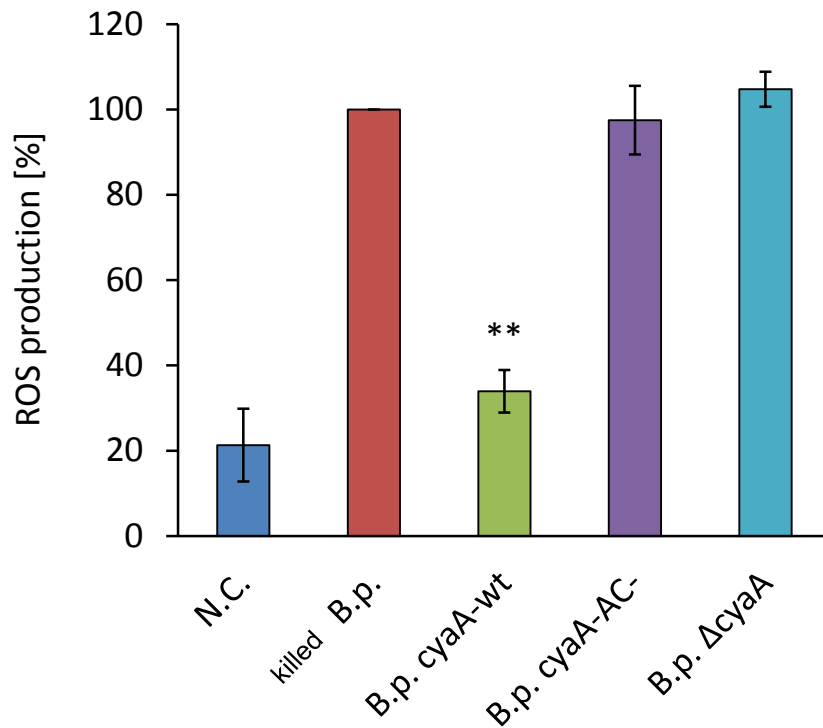


# cAMP-elevating AC activity of ACT (CyaA) enables *B. pertussis* growth in the presence of neutrophils



Cerny et al., unpublished

# AC activity of *B. pertussis* ACT blocks ROS production by neutrophils and NET formation



**Cerny et al. unpublished**

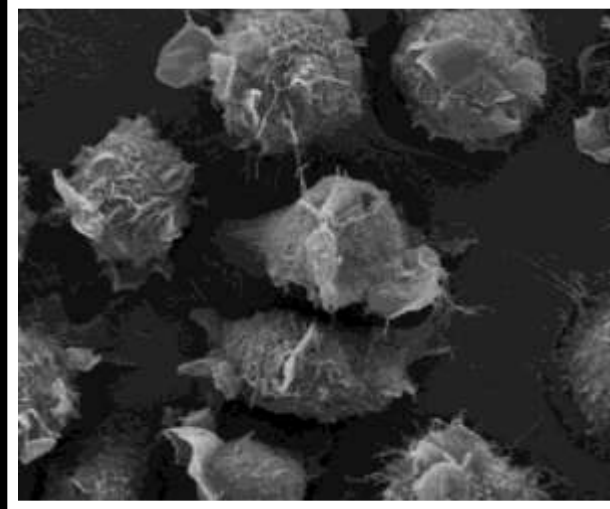
**Eby JC, Gray MC, Hewlett EL (2014). Infect Immun. 82:5256-69.**

- **cAMP-mediated suppression of neutrophil extracellular trap formation and apoptosis by the *B. pertussis* adenylate cyclase toxin.**
- **Convalescent-phase antisera from patients recovering from pertussis block ACT-mediated inhibition of the oxidative burst and NET formation**

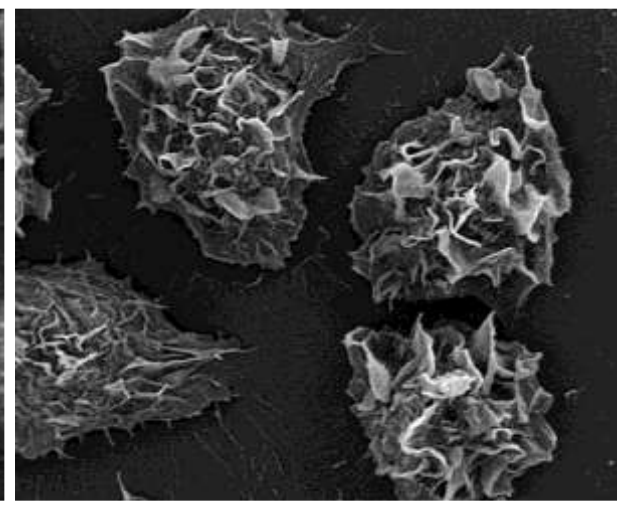
# CyaA-induced morphological rearrangements

Mouse  
macrophage-like  
cell line  
J774 A.1:

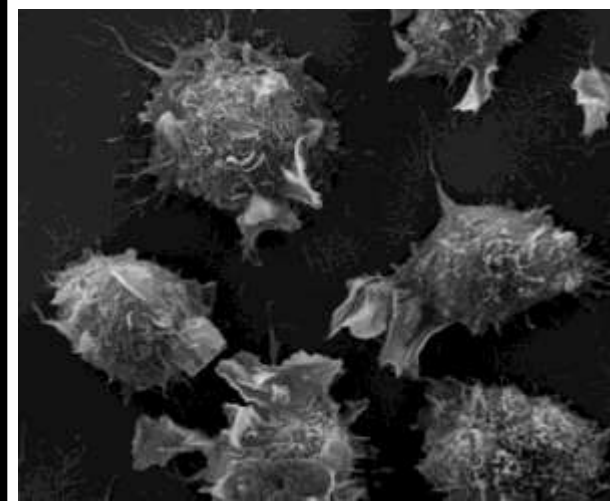
Buffer, 5 min



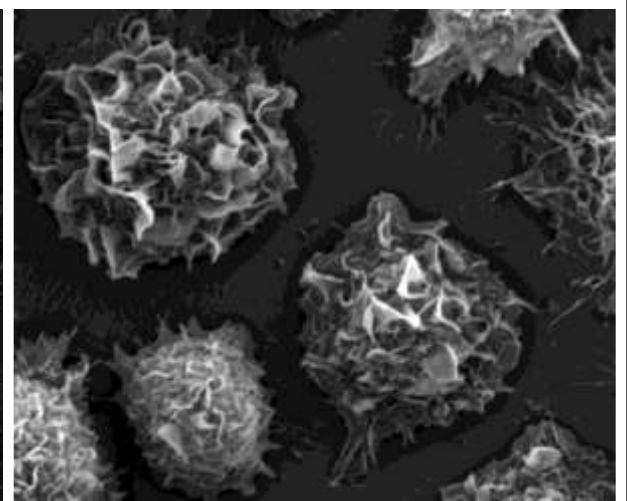
CyaA, **10 ng/ml**, 5 min



CyaA-AC<sup>-</sup>, 10 ng/ml, 5 min



db-cAMP, 2mM, 10 min

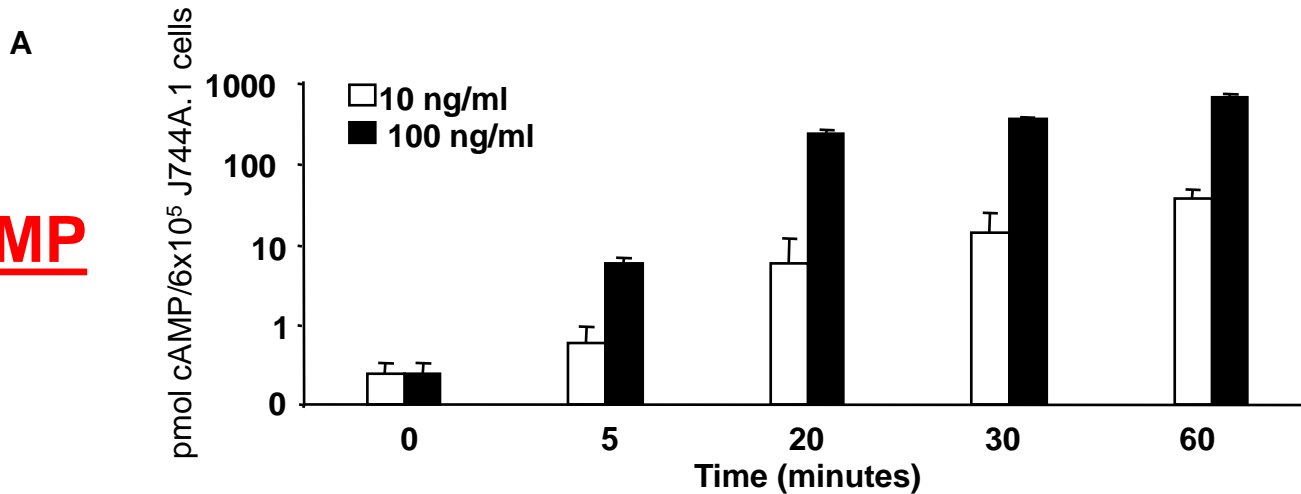


JanaK

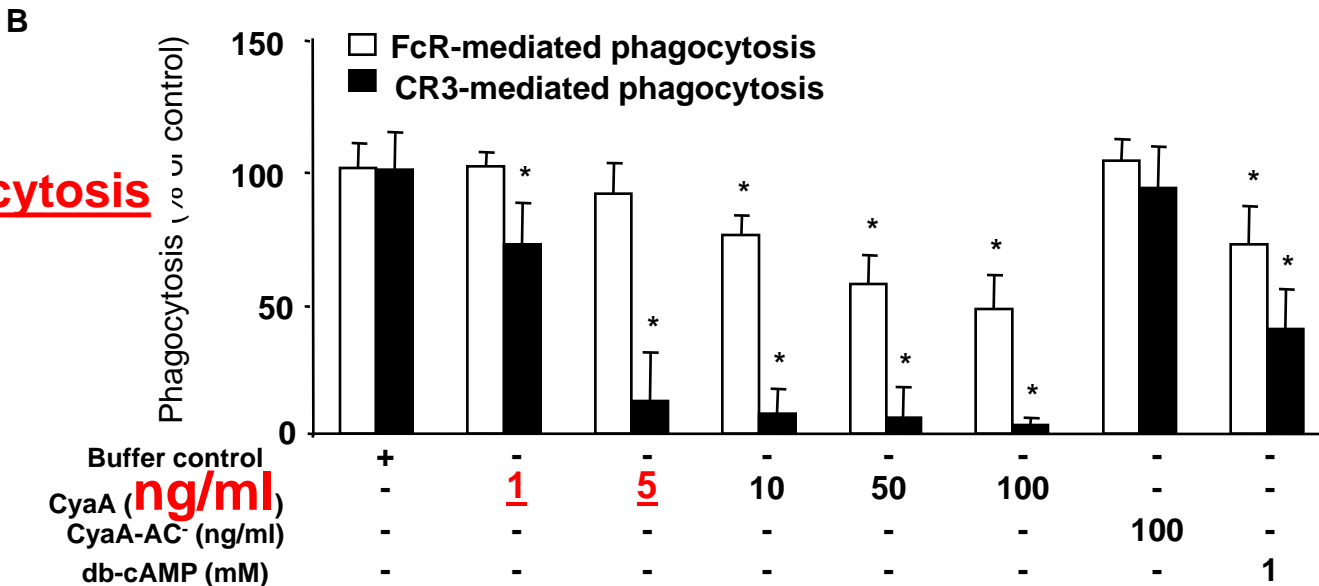
**Kamanova et al. (2008)**  
***J. Immunol.* 181, 5587-97**

# ACT at low doses ablates complement-mediated opsonophagocytosis (through RhoA inactivation)

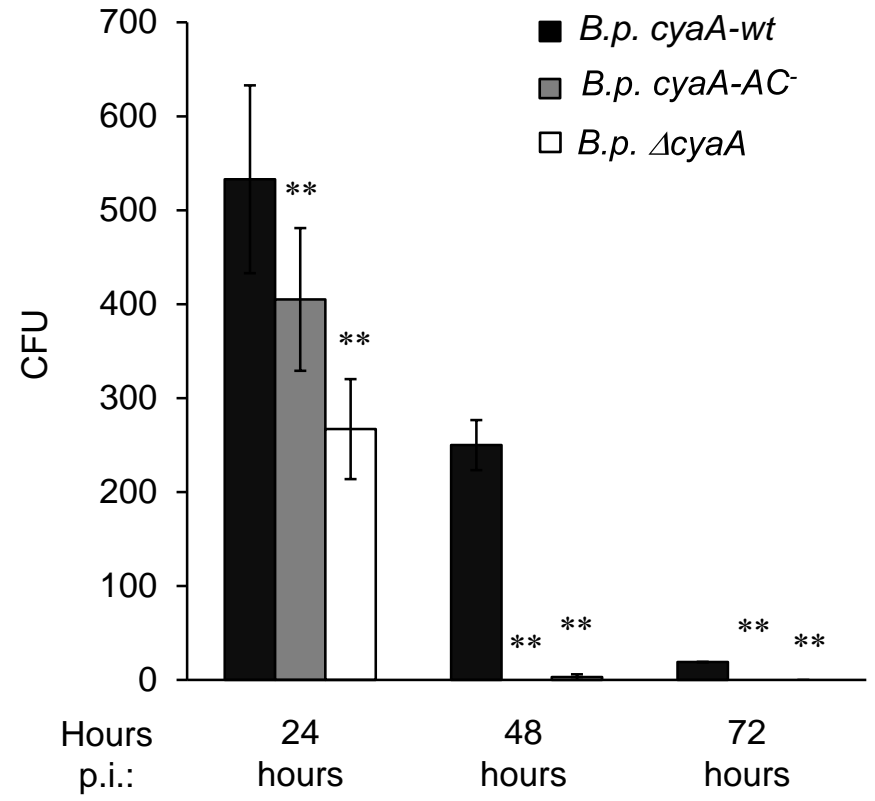
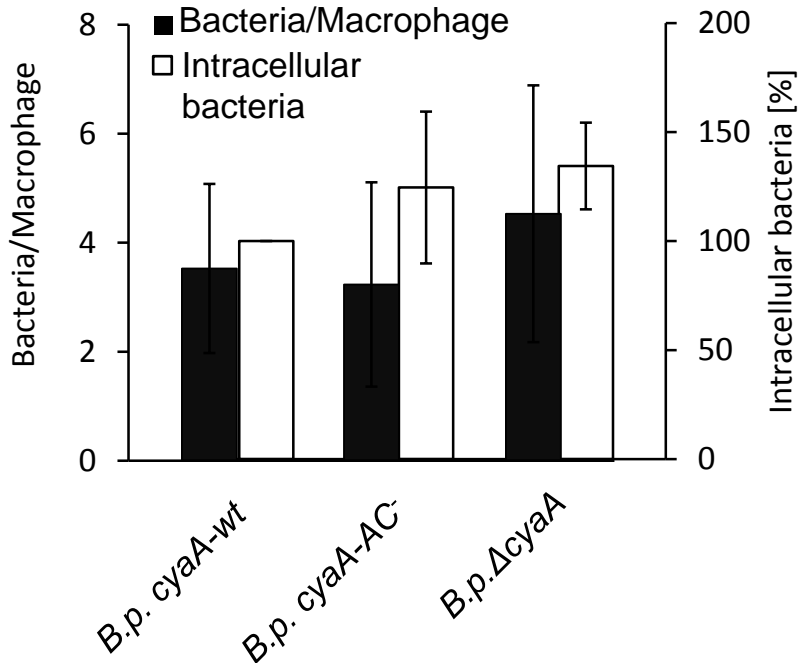
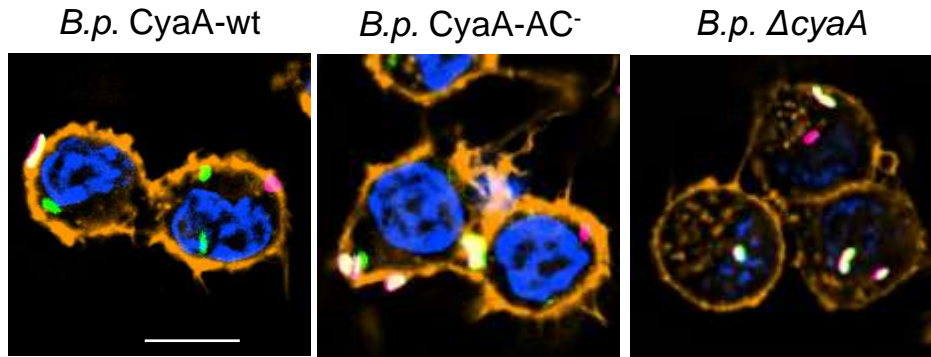
**cAMP**



**phagocytosis**

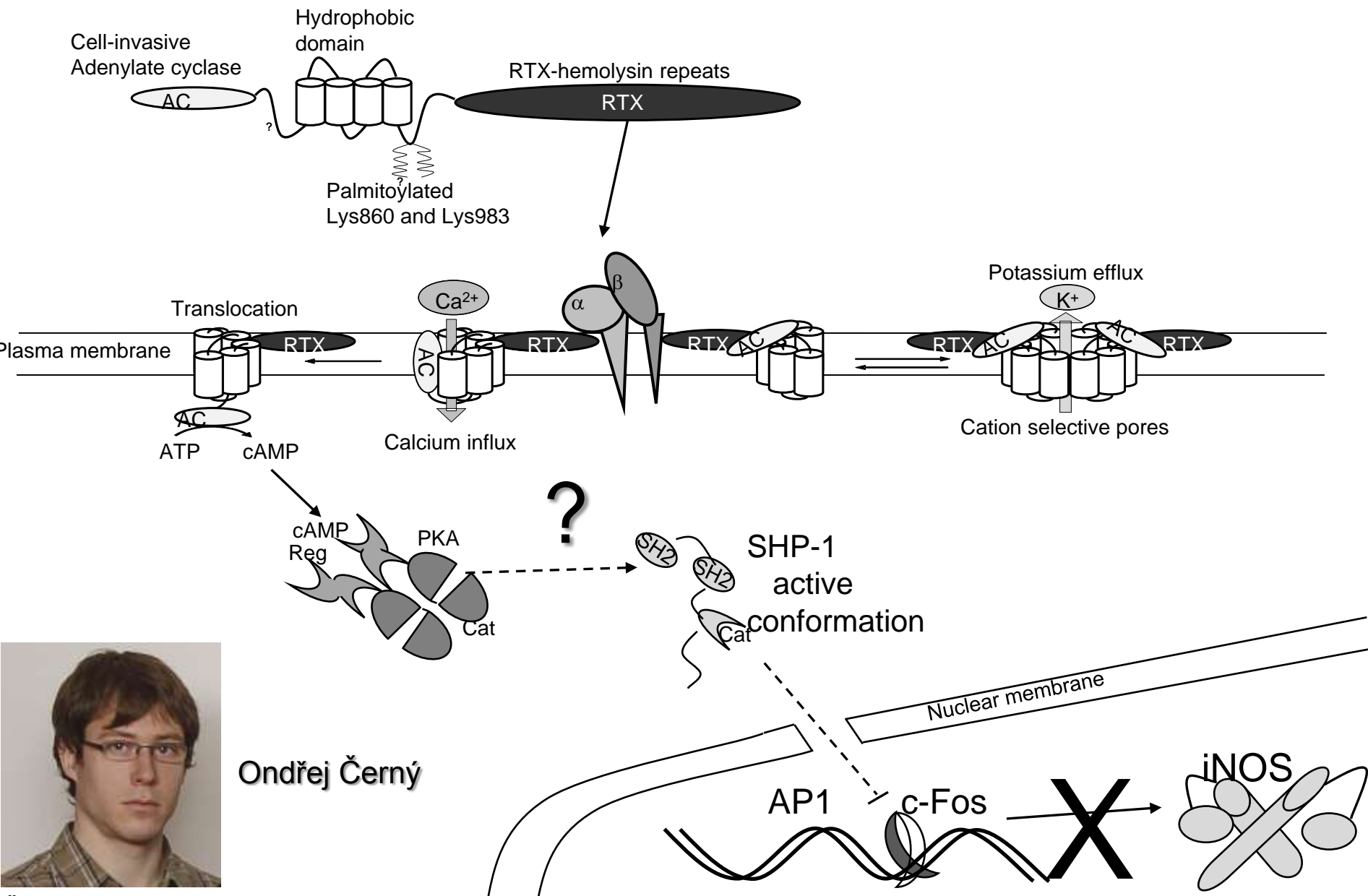


# Non-opsonized *B. pertussis* can persist within macrophages due to cAMP elevating activity of ACT





# ACT-produced cAMP signaling through PKA activates SHP-1 tyrosine phosphatase that blocks iNOS gene transcription

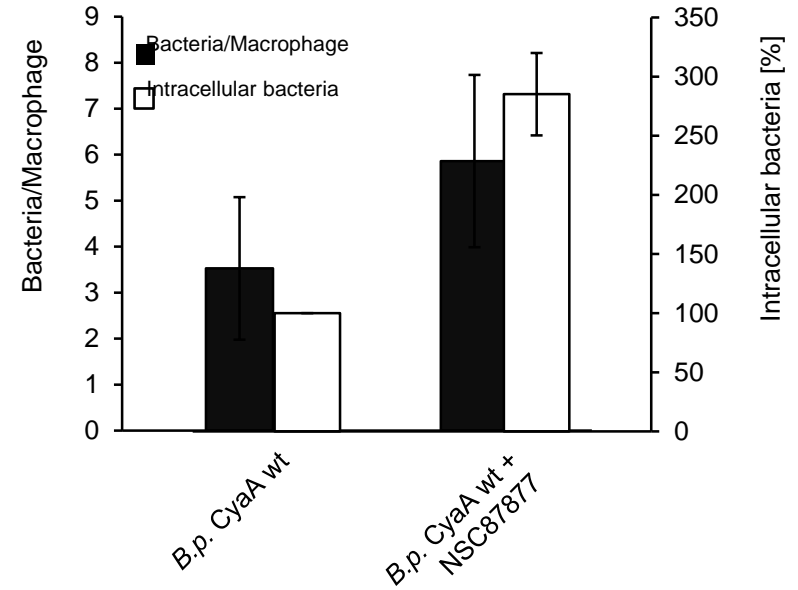
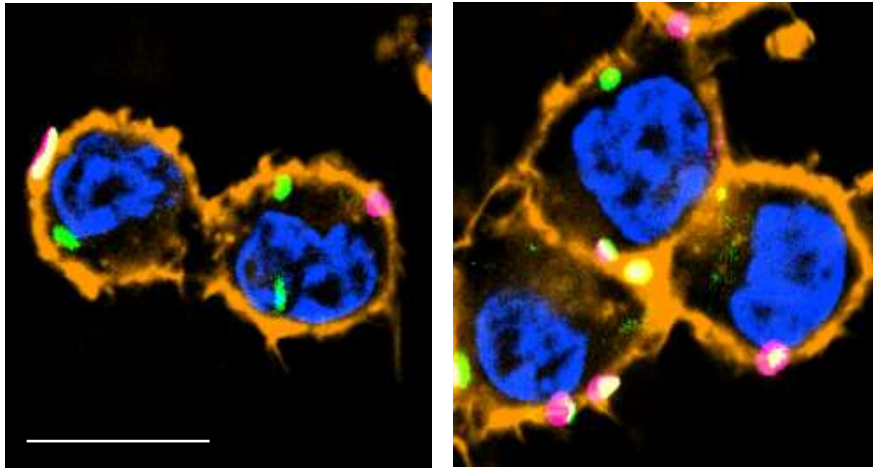


Ondřej Černý

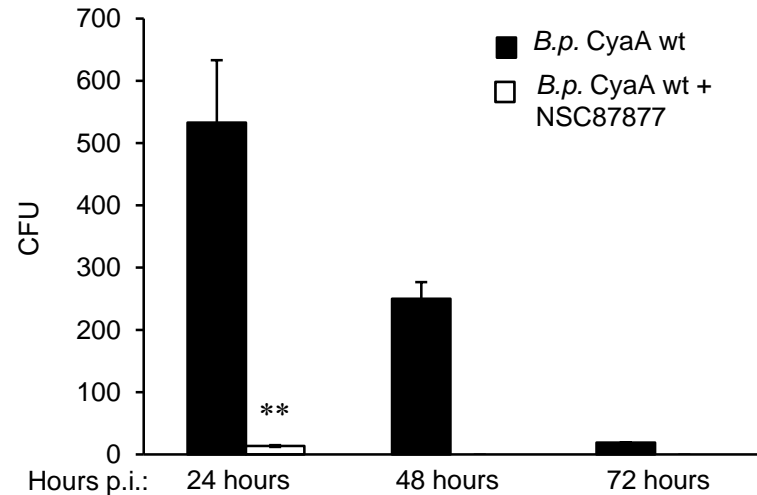
# CyaA/cAMP-triggered SHP-1 activation extends *B. pertussis* survival inside macrophages

*B.p.* CyaA wt

*B.p.* CyaA wt + NSC87877

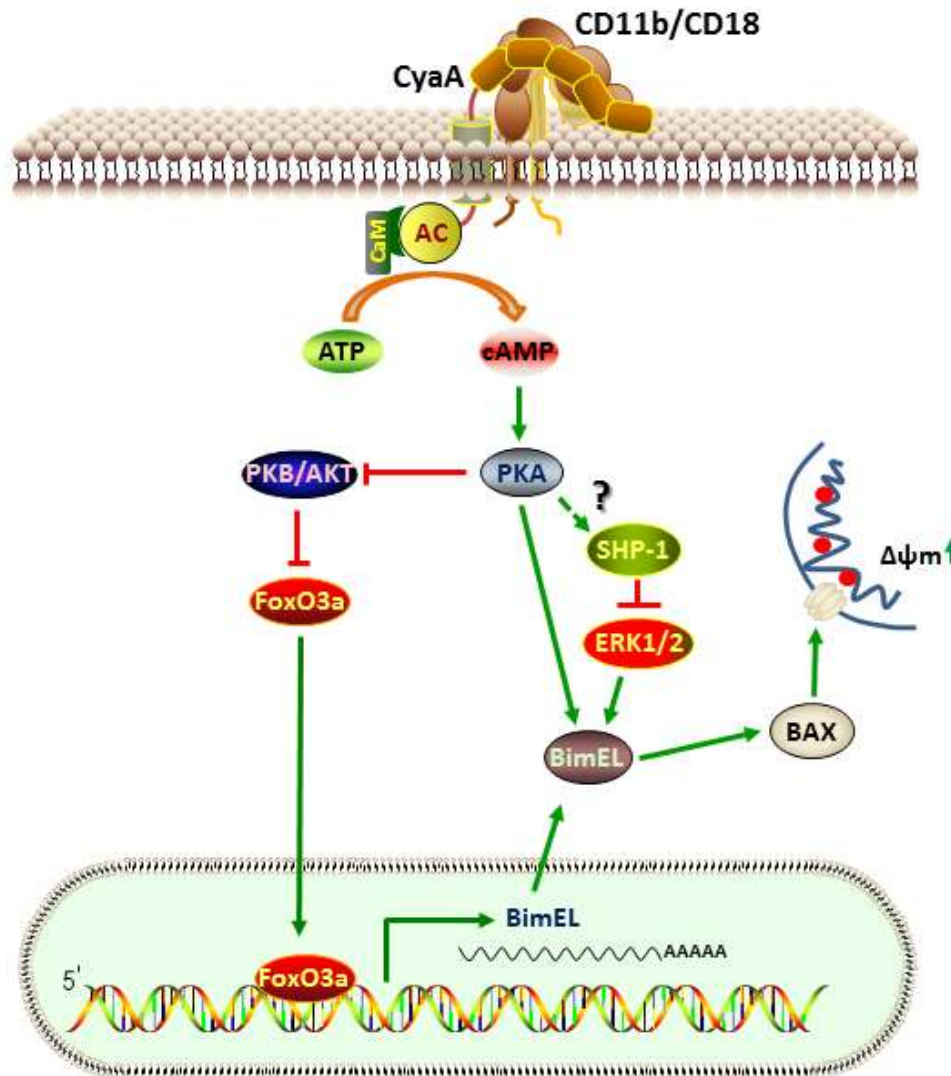


Ondra

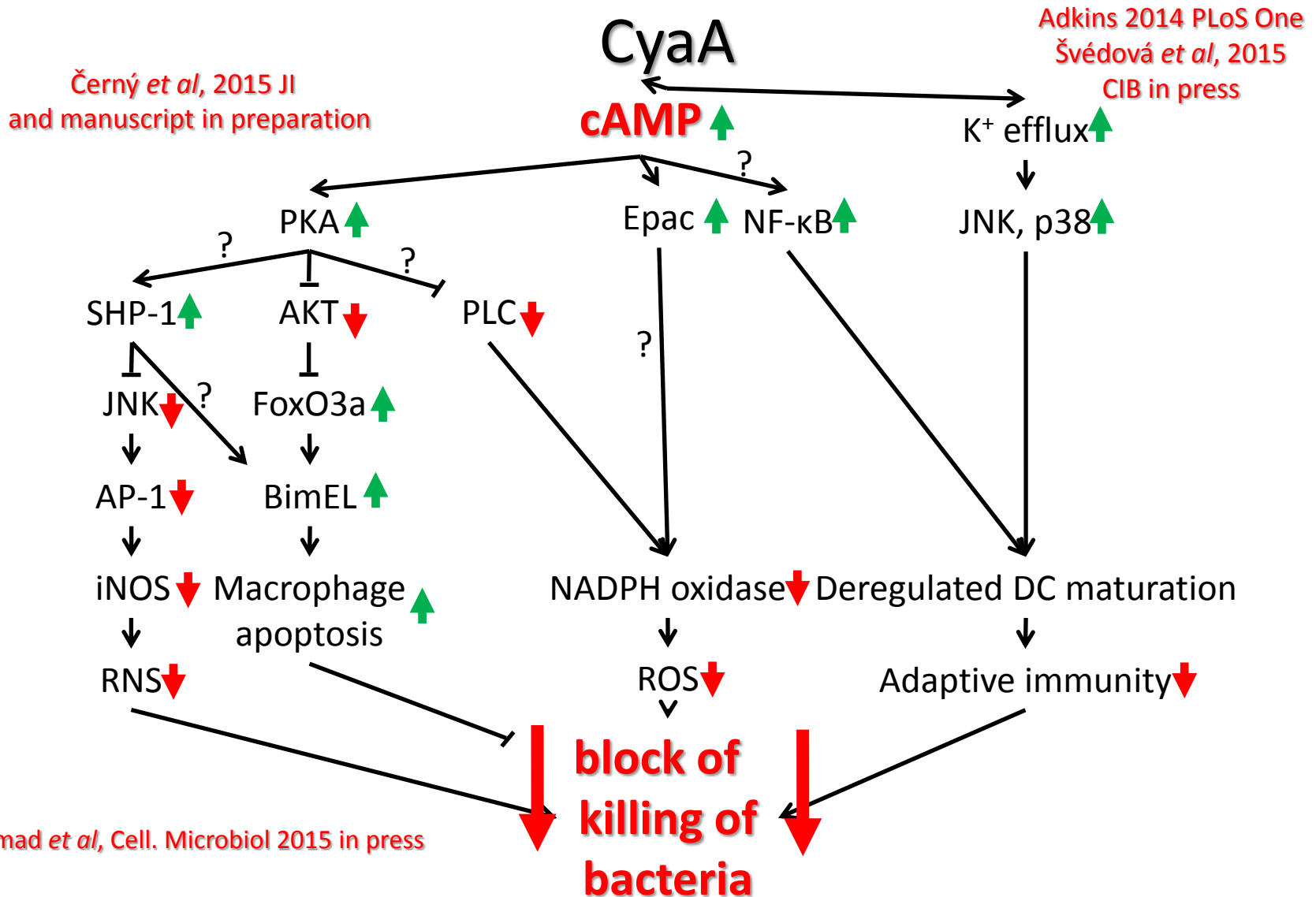


# CyaA/cAMP-triggered SHP-1 activation causes BimEL stabilization, Bax activation and macrophage apoptosis

Ahmad et al. 2015, Cell. Microbiol, in press

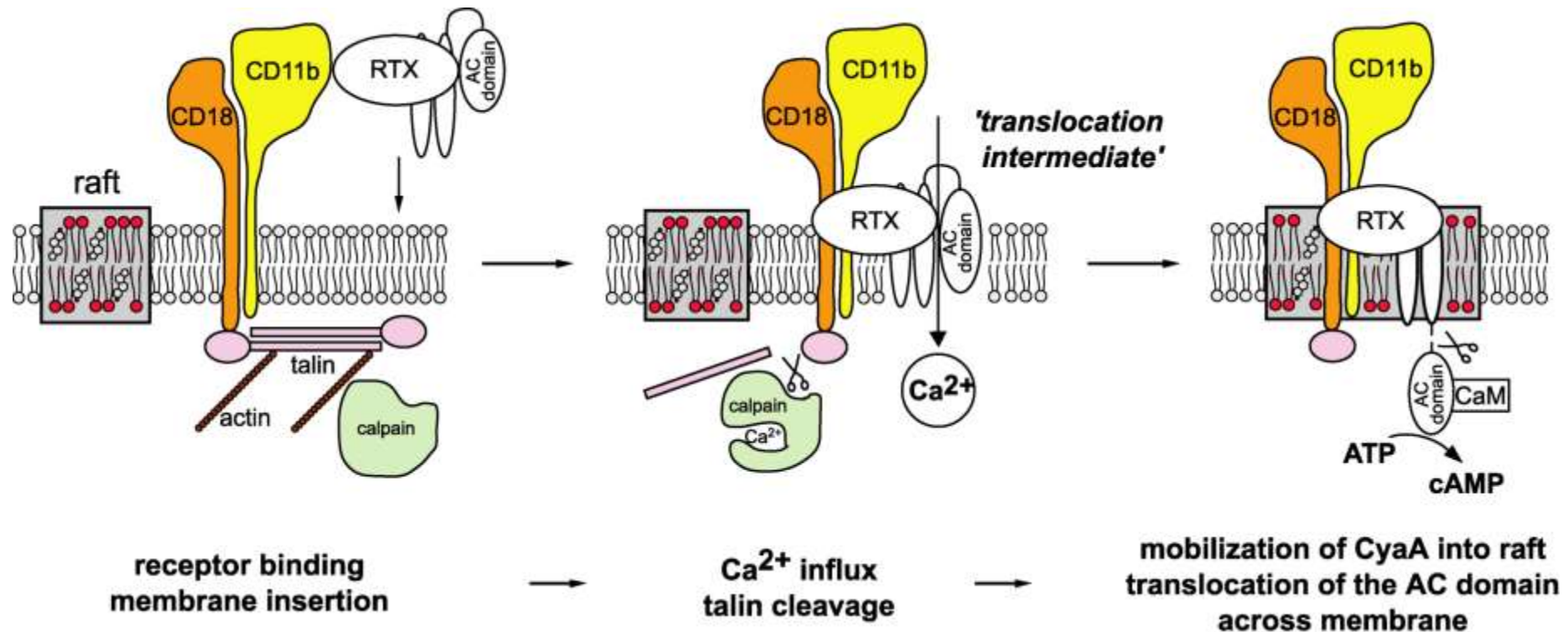


# Signalling pathways influenced by CyaA



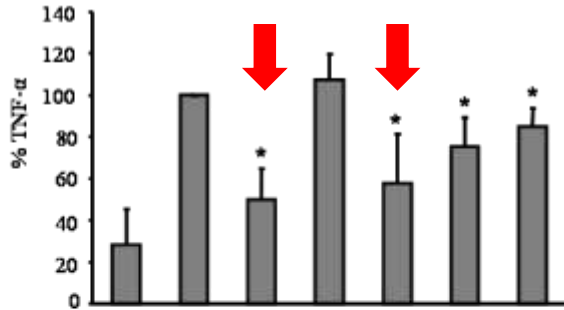
# Suppressing adaptive immunity

# Adenylate cyclase toxin hijacks the $\beta_2$ integrin receptor into lipid rafts to accomplish membrane translocation in two steps

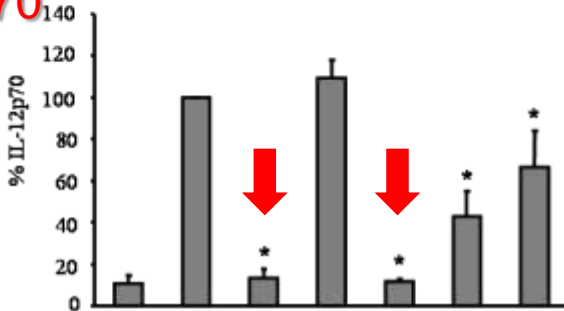


Bumba et al. (2010). PLoS Pathog 6(5): e1000901.

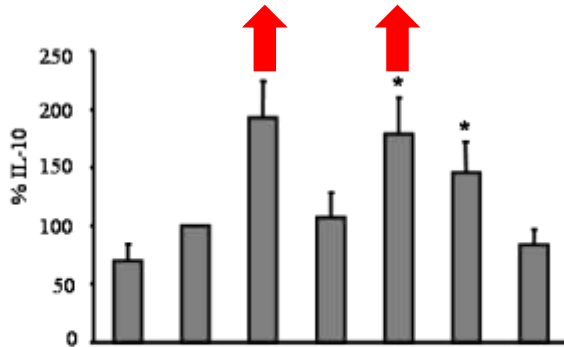
TNF $\alpha$



IL-12p70



IL-10



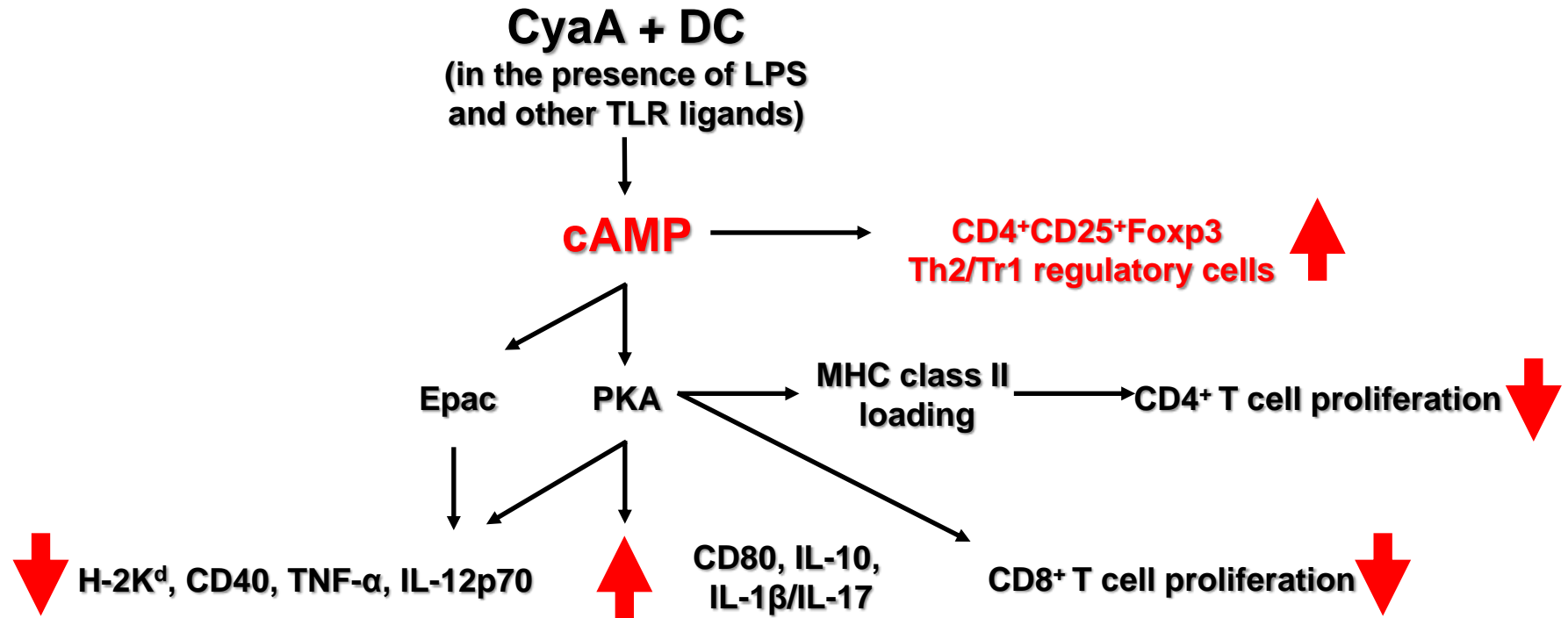
DC w/o LPS	+	-	-	-	-	-	-
buffer	-	+	-	-	-	-	-
CyaA	-	-	+	-	-	-	-
CyaA-AC	-	-	-	+	-	-	-
db-cAMP	-	-	-	-	+	-	-
6-Bnz-cAMP	-	-	-	-	-	+	-
8-pCPT-cAMP	-	-	-	-	-	-	+

ACT (CyaA) skews  
TLR-stimulated  
cytokine production  
in DC  
towards tolerance?

n = 4, \* P < 0,05; 100 % urea + LPS (buffer)

Irena Adkins

# cAMP signaling of ACT dampens and skews adaptive immunity towards initial $T_{h2}/T_{r1}$ -mediated tolerance of colonizing *Bordetellae* followed by delayed $T_{h1}/T_{h17}$ mediated clearance?



## Several reports indicated that CyaA subverts adaptive T cell immune responses

<i>Relman lab:</i>	Boschwitz <i>et al.</i> (1997) <i>JID</i> 176:678	Human MoDCs
<i>Guiso lab:</i>	Njamkempo <i>et al.</i> (2000) <i>J. Cell. Physiol.</i> 183:91	Human Monocytes
<i>Lewis lab:</i>	Bagley <i>et al.</i> (2002): <i>J. Leukoc. Biol.</i> 72:962	Human MoDCs
<i>Mills lab:</i>	Ross <i>et al.</i> (2004) <i>Infect. Immun.</i> 72:1568	Mouse BMDCs
<i>Mills lab:</i>	Boyd <i>et al.</i> (2005) <i>J. Immunol.</i> 175: 730	Mouse BMDCs
<i>Ausiello lab:</i>	Spensieri <i>et al.</i> (2006) <i>Infect. Immun.</i> 74:2831	Human MoDC
<i>Mills lab:</i>	Hickey <i>et al.</i> (2008) <i>J. Leukoc. Biol.</i> 84:234	Mouse BMDCs
<i>Ausiello lab:</i>	Fedele <i>et al.</i> (2010) <i>PLoS One.</i> 5(1): e8734	Human MoDC

[Adkins et al. \(2014\) PLoS One 9\(8\):e104064](#)



**Hence:**

The **cAMP** signaling of **ACT**,

prevails and

**knocks down the innate immunity**

and

**dampens the adaptive immune  
response**

in order

**to enable host colonization by  
*Bordetellae***

Explaining why is it so important  
to add the AC toxoid  
into the aP vaccine

if we are serious about

breaking the vicious circle  
of epidemic whooping cough  
spread in the most developed  
countries ...

## To sum up this talk:

Rise in pertussis incidence in the most developed countries will likely not stop until the adenylate cyclase toxoid has been added to the aP vaccine...

And a less reactogenic nextgen wP vaccine is used as booster...

# Sebolab = a PI Confederation...



**Radim Osička** – CR3 interactions and cAMP signaling of ACT



**Irena Adkins** – ACT and fooling od DC maturation and cAMP-mediated inhibition of T cell immune response



**Jiří Mašín** – membrane penetration and pore-forming activity of ACT



**Peter Šebo** – strategic steering, mentoring and paper editing, signaling of cAMP and phagocyte action



**Láďa Bumba** – structure, folding and secretion of ACT



**Ondřej Staněk**  
novel antigen delivery tools  
Veterinary vaccines



**Branko Večerek** – sRNA regulation of *Bordetella virulence*



**Jana Holubová + Karolína Škopová**  
novel pertussis whole cell vaccine

# External collaborations:

## Institut Pasteur:

teams:

**Nicole Guiso**

**Claude Leclerc**

## University of Virginia

**Erik L. Hewlett**

## Trinity College

**Kingston Mills**

Aisling Dunne

## Institute of Microbiology

Lída Tučková

Marek Kovář

and their teams

## University Wurzburg

Roland Benz

and his team

## Bernhard Nocht Institut:

Thomas Jacobs

Susanne Tartz

## MH Hannover

Ingo Just

Harald Genth

## VLA Surrey

Martin Vordemeier