



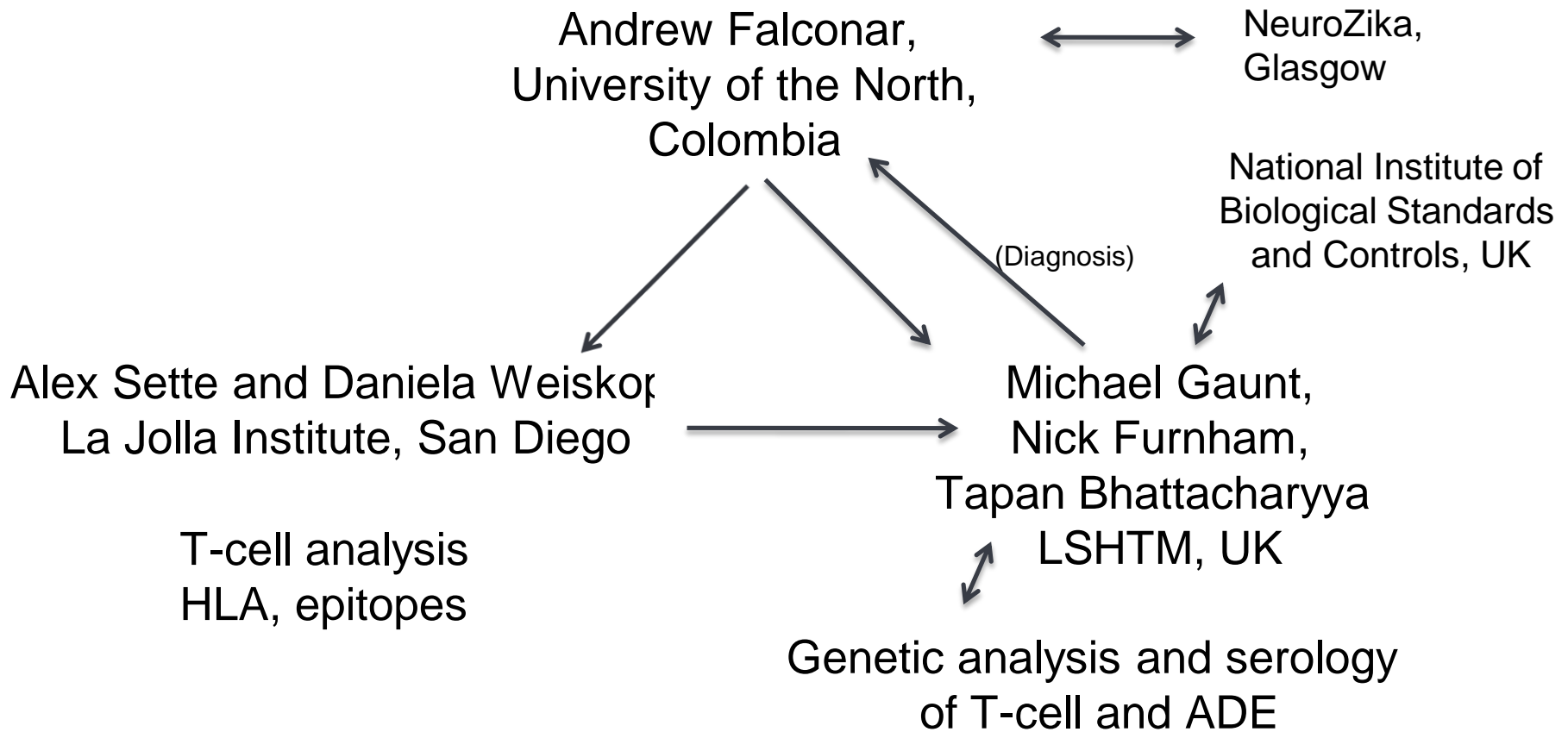
# INVADE: Investigating Antibody Dependent Enhancement Finding the mutations

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Weiskopf and Alexander Sette

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partner authoring  
the document here

12/10/2016

# General setup: WP 6, INVADE

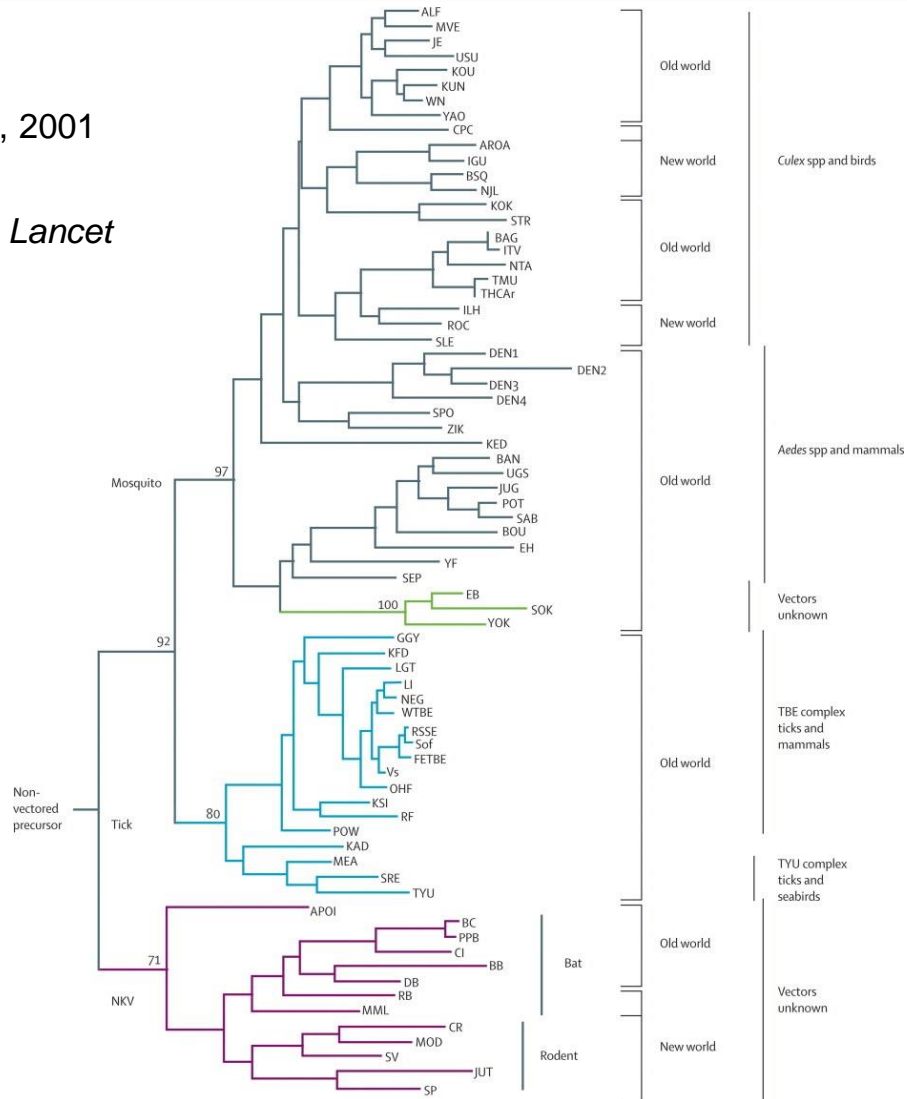




# Zika virus and Evolution

Gaunt *et al.* 2001, reviewed Ash, 2001  
*Science*

Lancet Gould and Solomon 2008 *Lancet*  
371, 500–509,  
+ 8 additional publications





# Molecular basis to Butantan vaccine

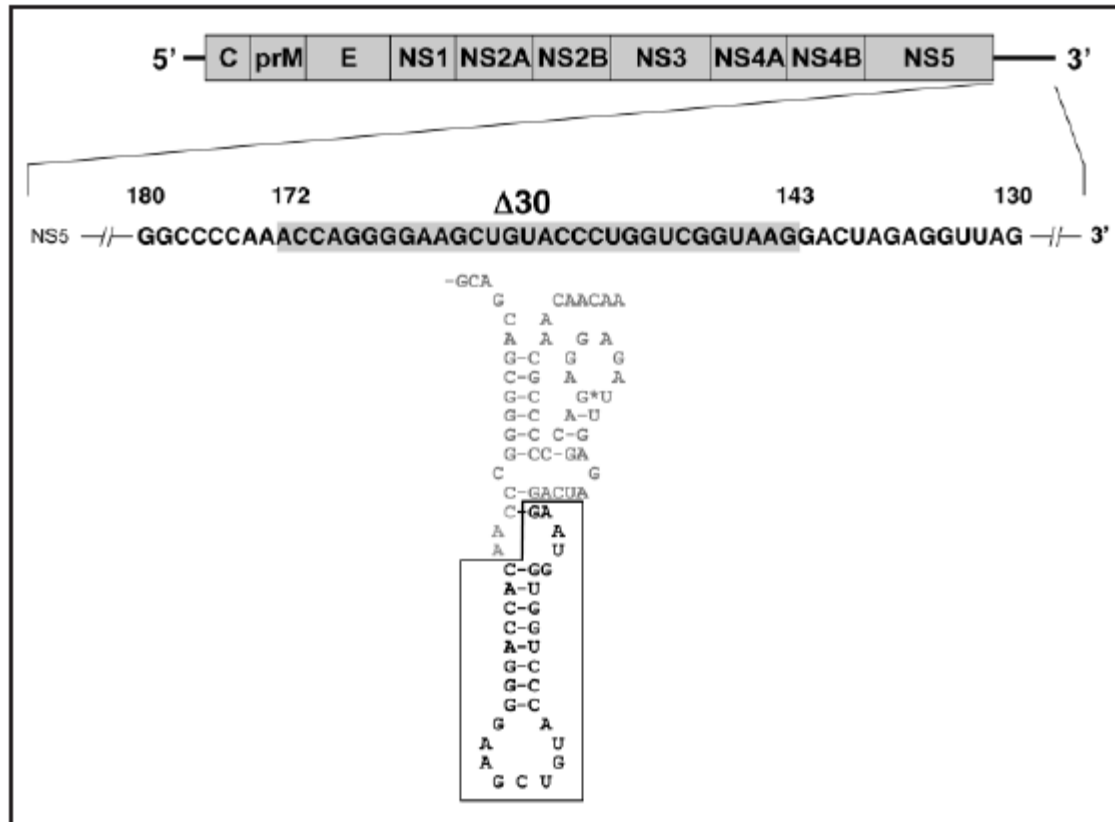


Figure 1. The  $\Delta 30$  mutation is located in the 3'UTR region of the DEN1 genome. The highlighted nucleotides represent those included in the deletion. Below the sequence is the predicted secondary structure of the TL2 region of DEN1 with the deleted nucleotides in the highlighted sequence boxed and in bolder type.

Copyright  
<http://weallseqtoseq.blogspot.de/2013/09/niaid-dvi-tv003-live-attenuated.html>

Whitehead *et al.* (2007)





# Molecular basis to Butantan vaccine

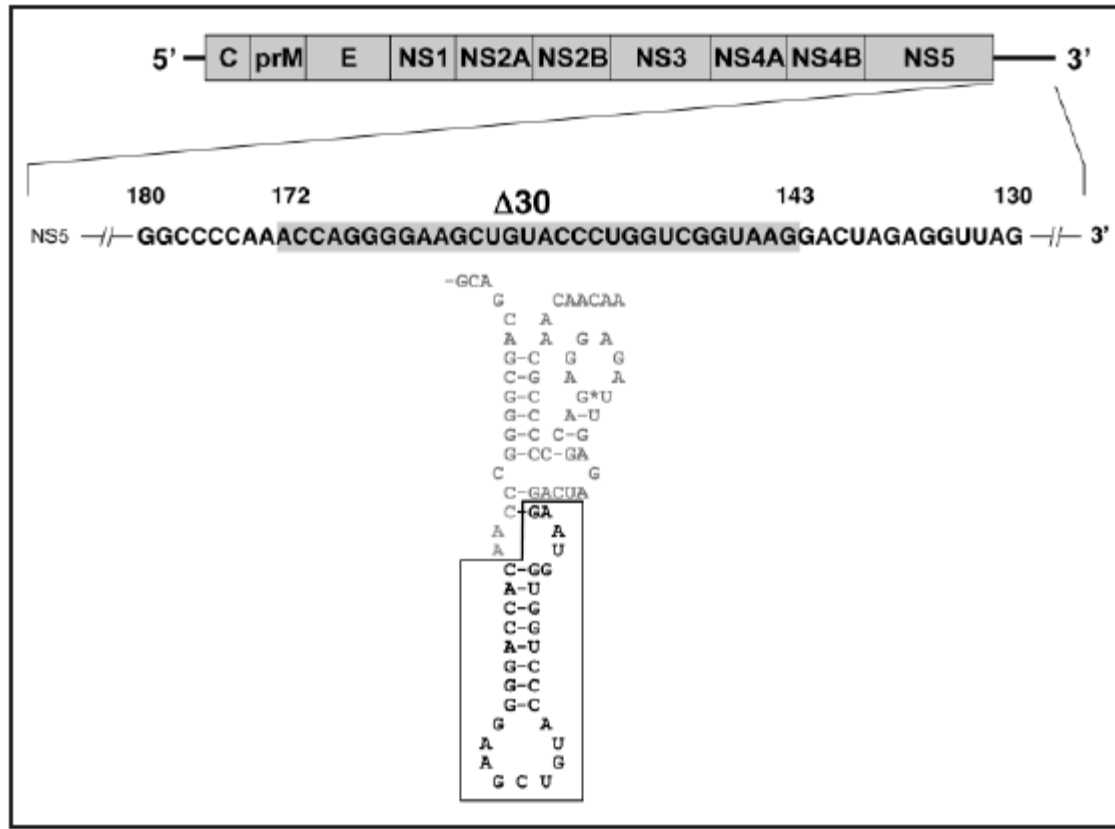


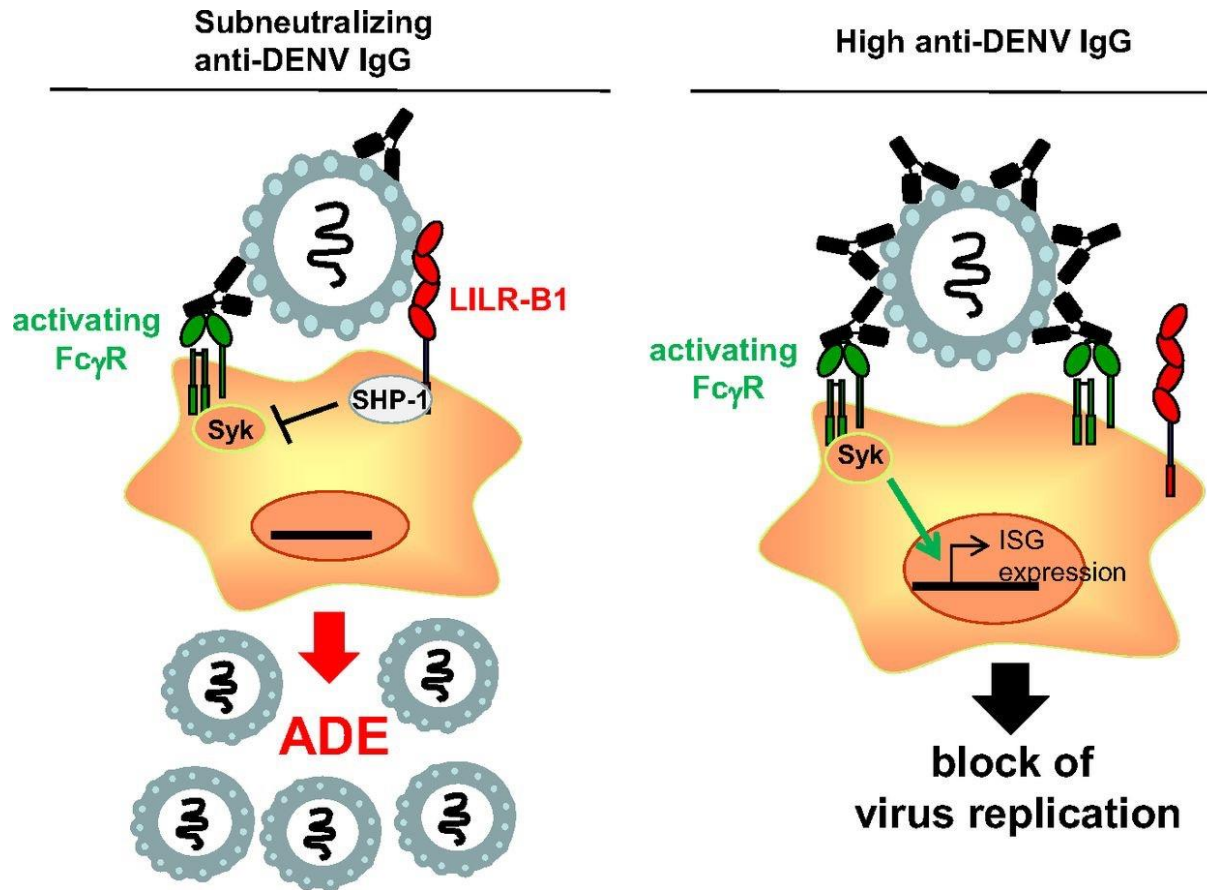
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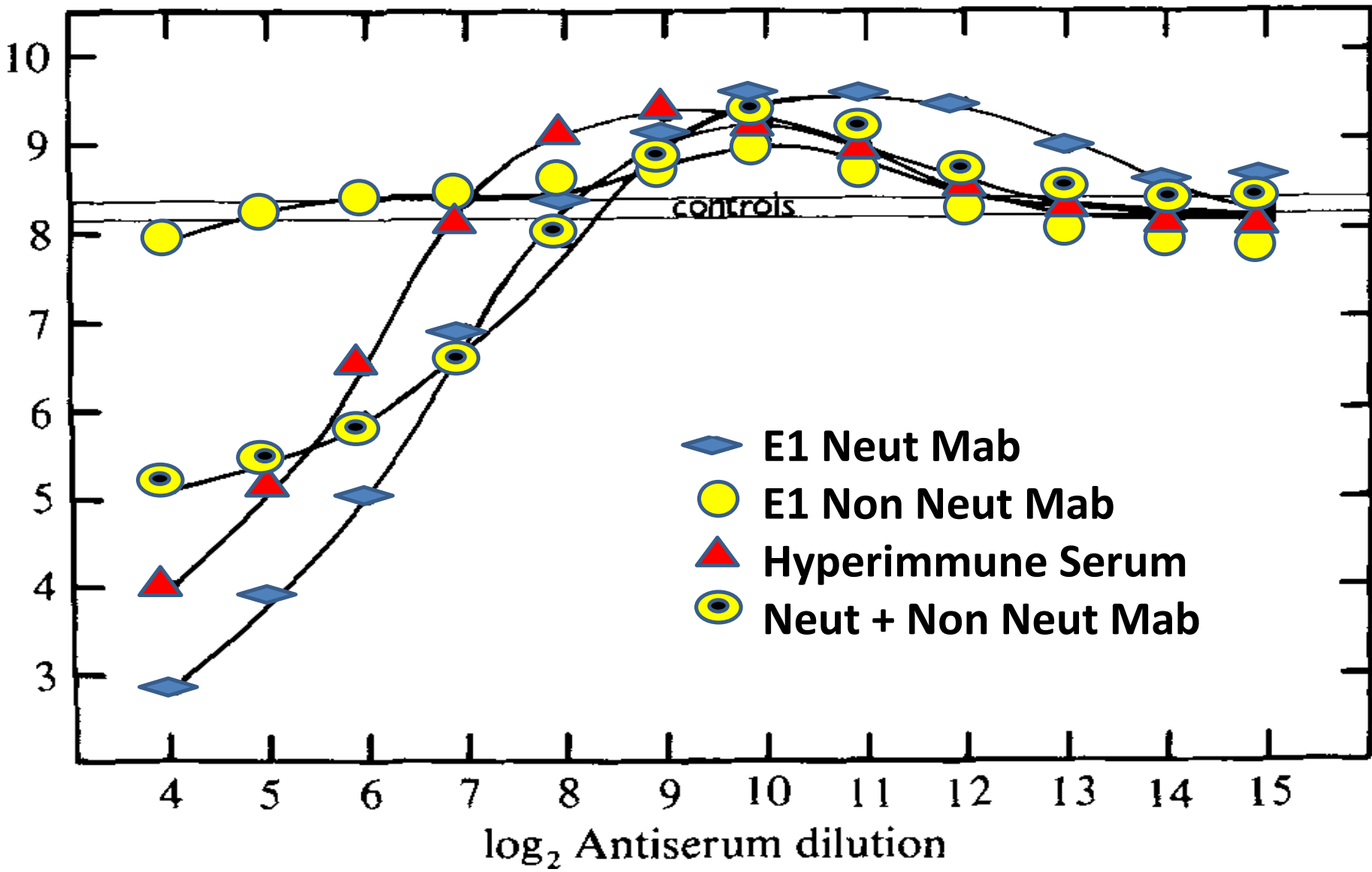


# Antibody dependent enhancement *in vitro*

First described by Halstead  
and developed as an *in vitro* test by Peiris and Porterfield



log<sub>5</sub> p.f.u./ml



Antibody dependent enhancement P388D1 cells (mouse macrophage)

# ADE Macaque Study

Four monkeys challenged with DENV1 or DENV2 2.8 years before Zika virus challenge

Four controls

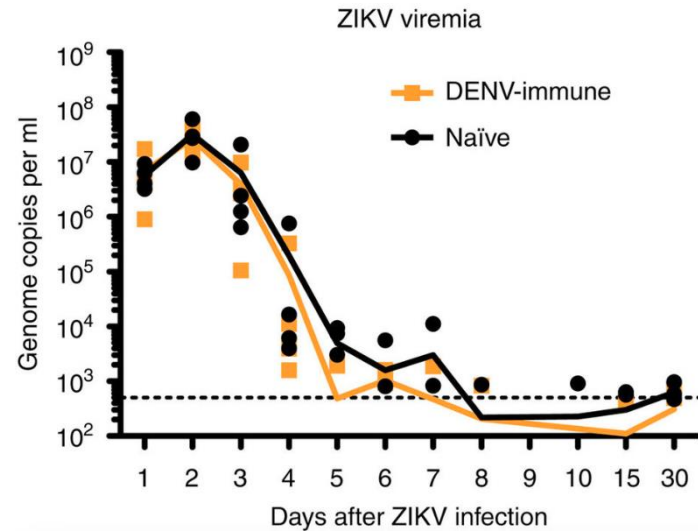
Pantoja *et al.* (2017)

Nature Comm. 8: 15674

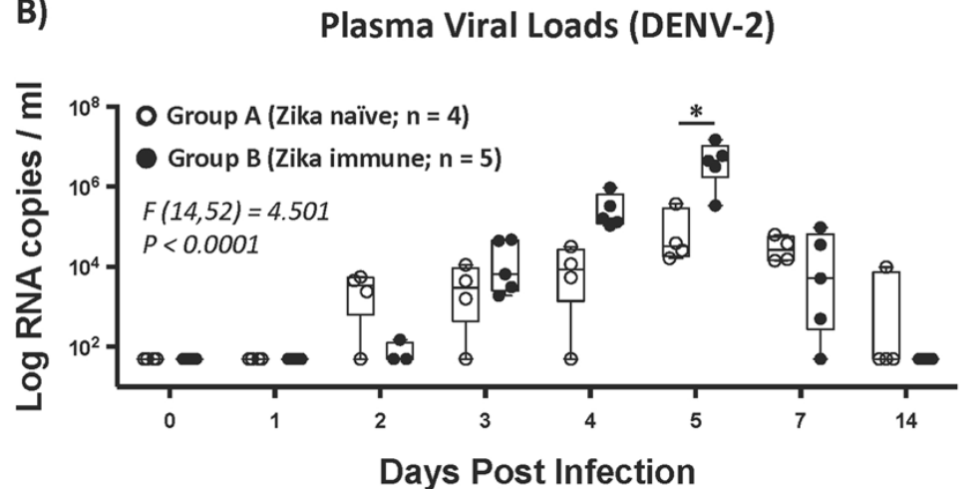
Five monkeys challenged with ZIKV before DENV2 challenge

George *et al.* (2017) Scientific

Reports. 8: 15674



B)







# Sample base to date

- 📍 650 ZIKV sera from humans ~400 RT-PCR for Zika virus from Caribbean (400ul)
- 📍 ~50 DENV IgM sera from the Caribbean
- 📍 Control ZIKV sera is available from travel patients without exposure to DENV
- 📍 All samples have relevant epidemiological information for age, gender and location
- 📍 **TIGHTEN THE PHYSIOLOGICAL CONDITIONS OF THE ASSAY**

# Bioinformatics of attenuating mutations for ADE

Virus	Monoclonal antibody		
	294	427	126
YF-FNV	7.0 ± 0.3	5.2 ± 0.3	5.4 ± 0.3
YF-Asibi	7.7 ± 0.2	6.6 ± 0.2	6.8 ± 0.1
YF-B11	8.8 ± 0.5	6.0 ± 0.7	7.4 ± 0.1
YF-B1	7.6 ± 0.7	8.0 ± 0.2	7.6 ± 0.3
YF-B5	9.2 ± 0.7	P	P
YF-B7	9.3 ± 0.3	10.8 ± 0.5	10.3 ± 0.6
YF-B9	8.3 ± 0.3	8.6 ± 1.1	8.8 ± 0.6
YF-B15	8.4 ± 0.5	8.1 ± 0.3	NT
YF-17D-UK	12.3 ± 0.3	P	P
YF-17D-SA	8.4 ± 0.4	P	P
YF-17DD-Braz	9.5 ± 0.4	P	9.8 ± 0.4
YF-9026	10.2 ± 0.3	9.8 ± 0.6	NT
JE	5.4 ± 0.2	5.9 ± 0.3	5.9 ± 0.4

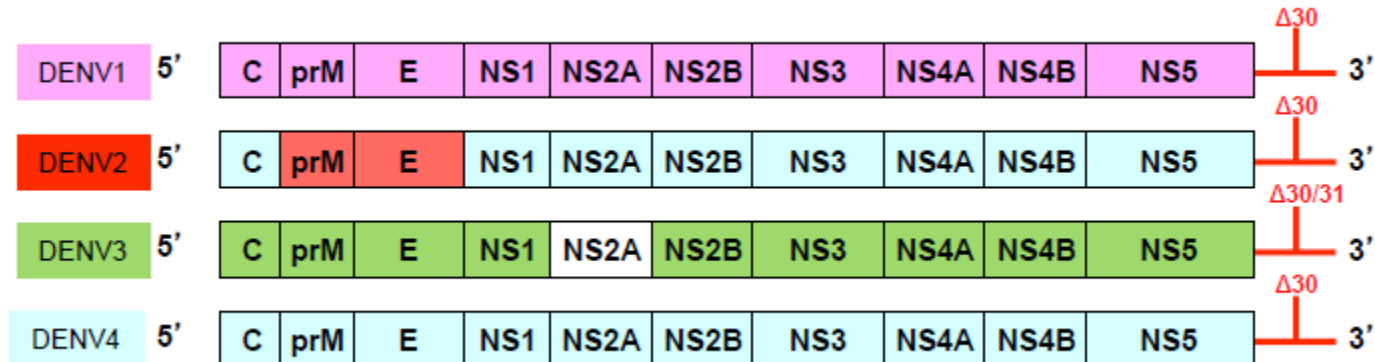


# Butantan vaccine: When modelling worked

- 📍 Tetravalent vaccine tested at the Butantan Institute, Brazil
- 📍 Attenuated mutations introduced in the 3' UTR
- 📍 Phase 3 clinical trials in Brazil
- 📍 Will induce a strong Th1 response

TV003

NIH National Institutes of Health  
Turning Discovery Into Health



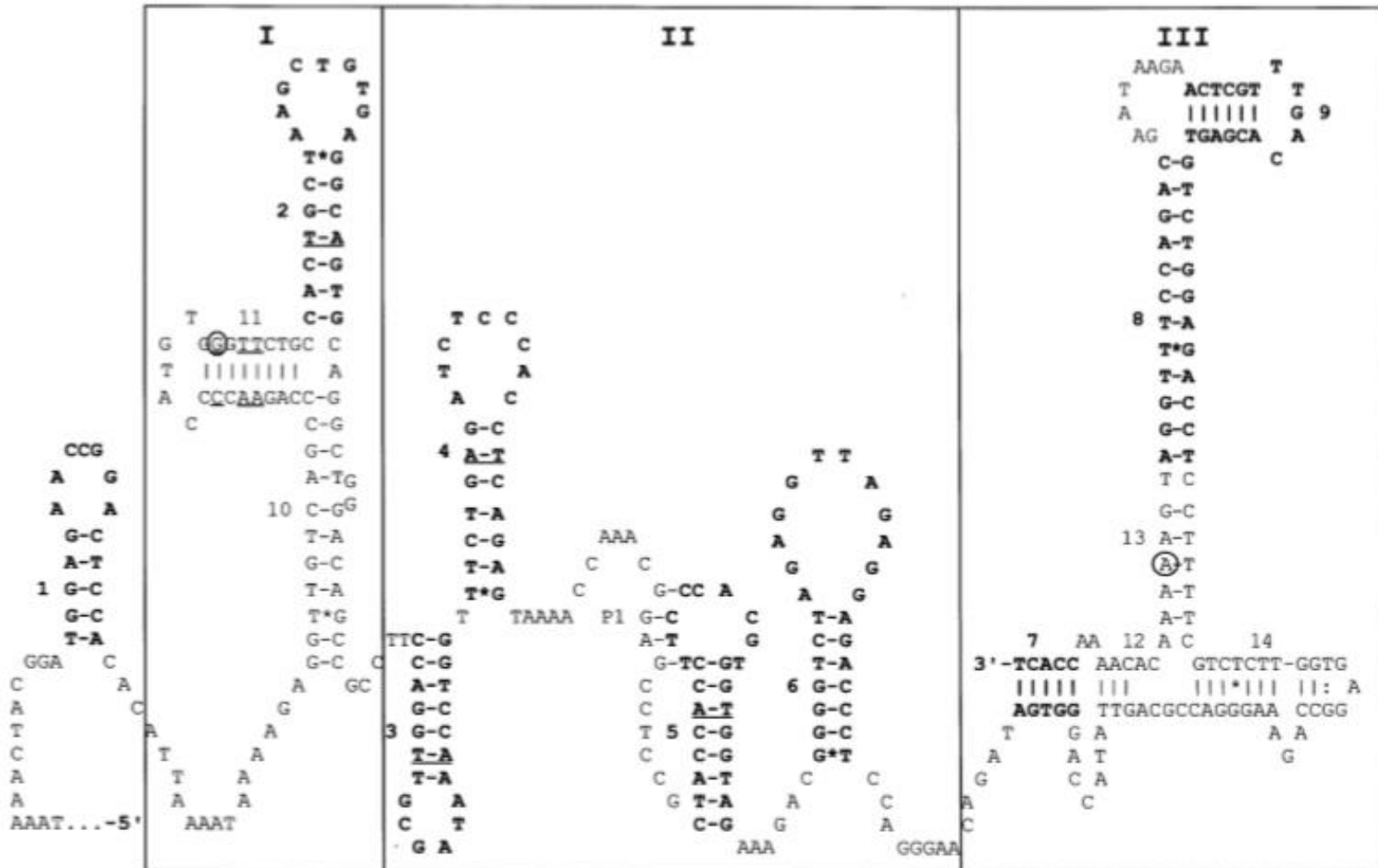
[http://1.bp.blogspot.com/-evIEFX2\\_7O4/UjJMLRM-sal/AAAAAAAAAClg/onkKKOCI6SI/s1600/Picture2.png](http://1.bp.blogspot.com/-evIEFX2_7O4/UjJMLRM-sal/AAAAAAAAAClg/onkKKOCI6SI/s1600/Picture2.png)

Whitehead *et al.* (2007)



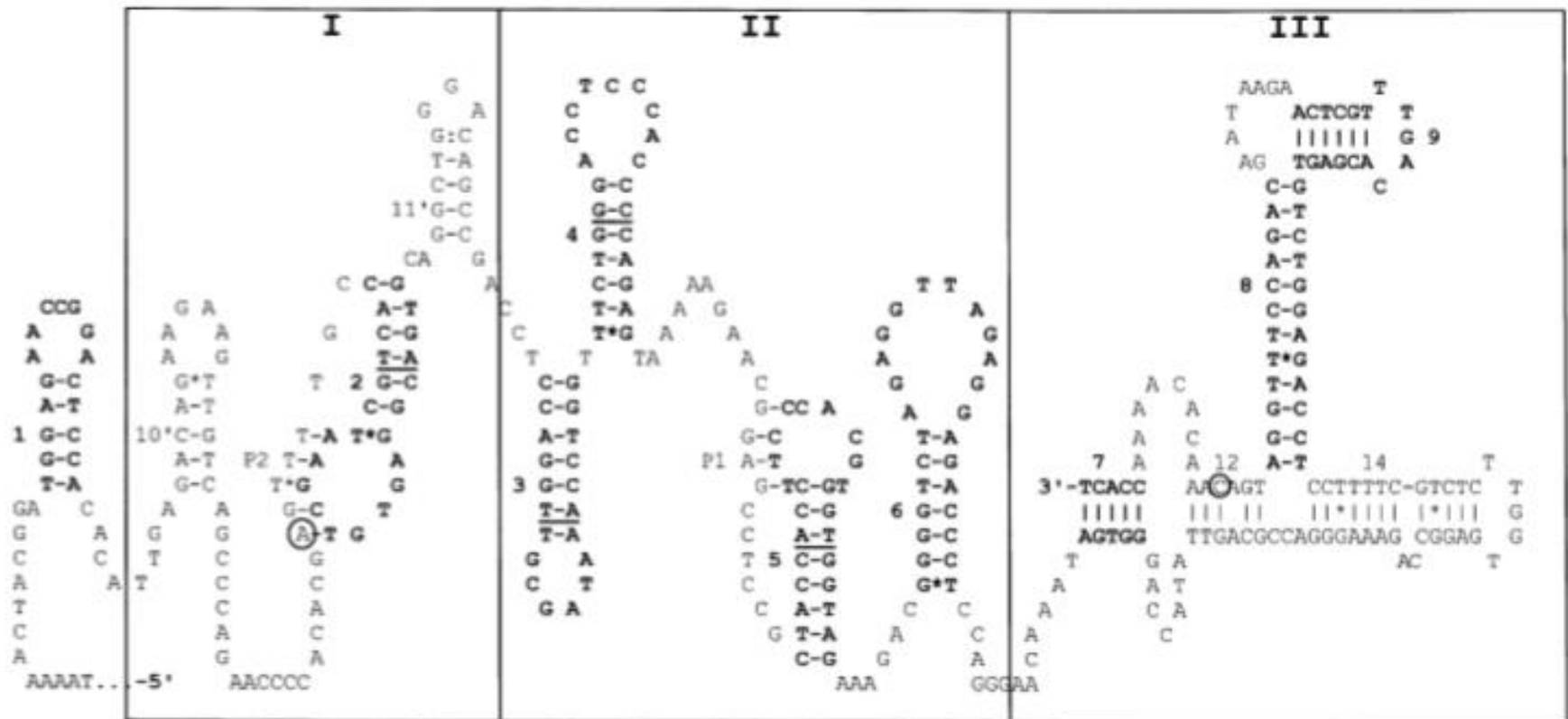
# When modelling worked

3' UTR secondary structure: wild type yellow fever virus



# When modelling worked

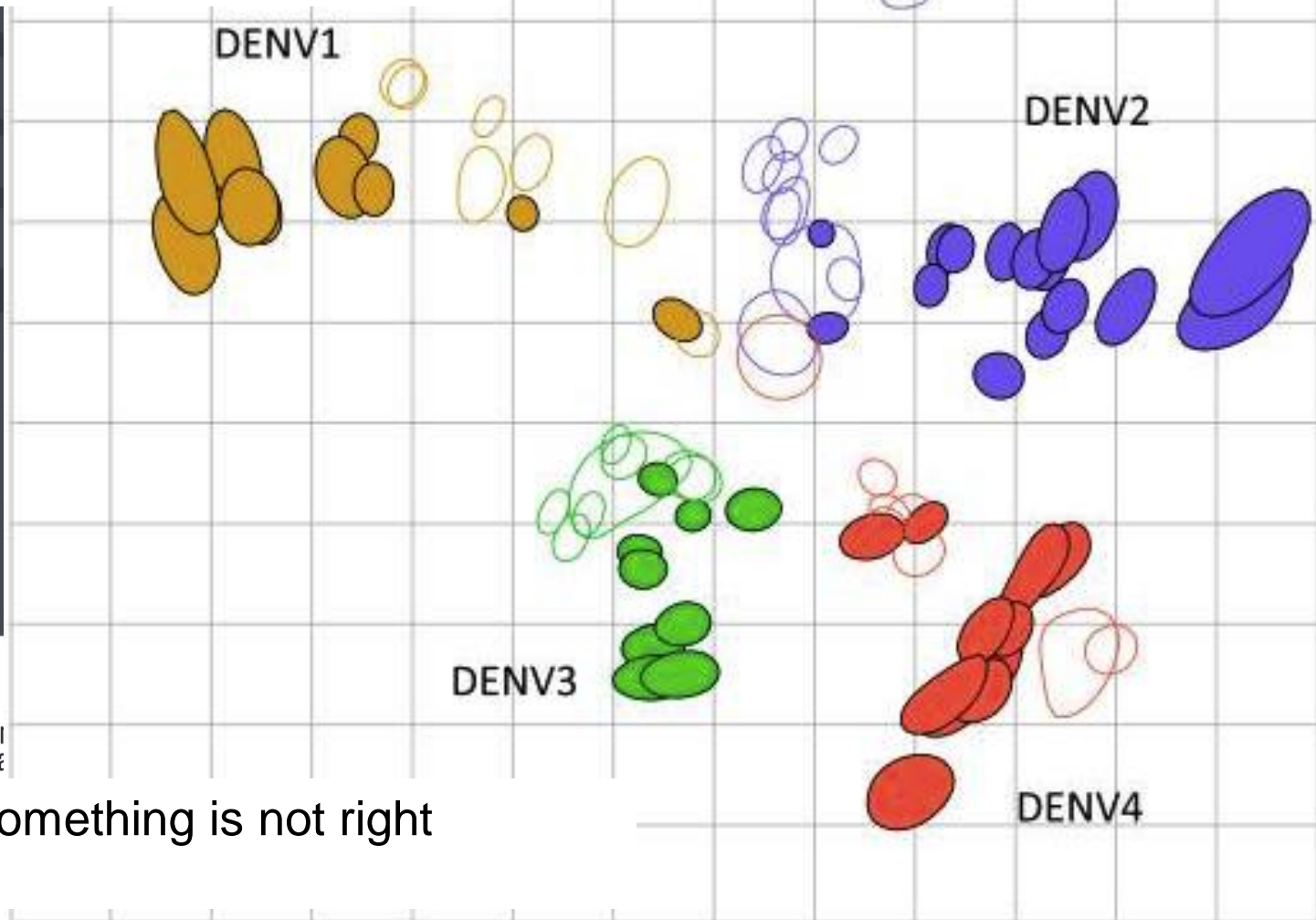
3' UTR secondary structure: vaccine yellow fever virus



# Antigenic map using PRNT for acute DENV sera

Serological analysis for cross-reactivity  
(PRNT) in a mouse model

KATZELNICK *et al* (2015) *Science* 349: 1338-1343

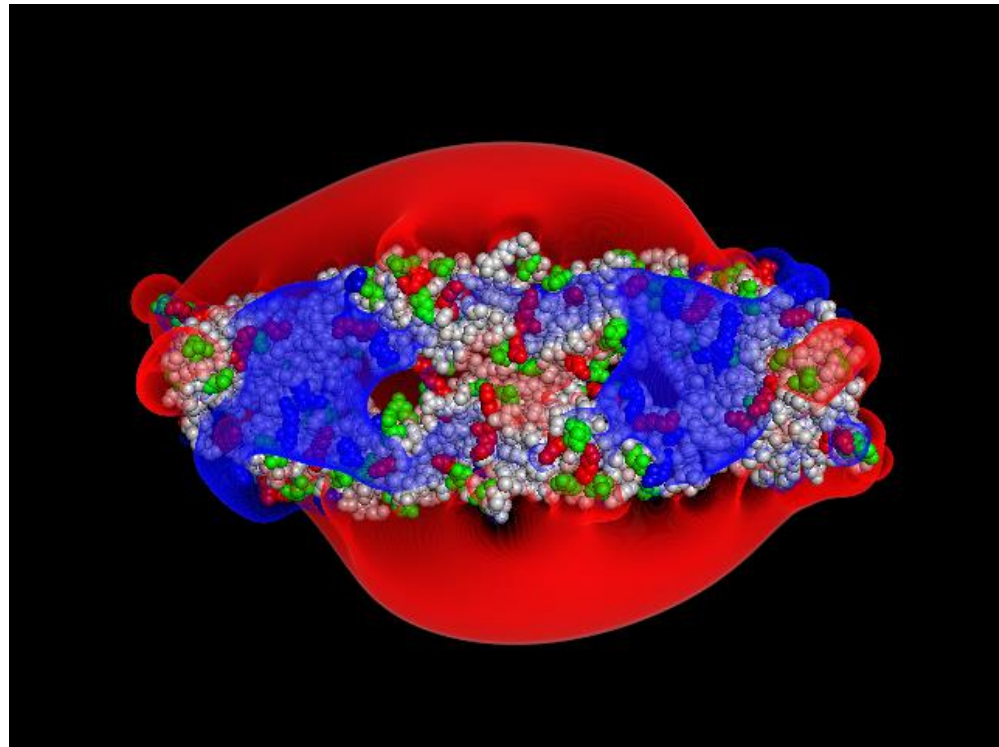


Something is not right

# What is the molecular mechanism of ADE?

## 📍 Hinge regions, or conformational plasticity?

- DENV E-protein: Domain II (fusion protein) relates to DI and DIII as a 'hinge' or hairpin



Work in collaboration with  
Nick Furnham  
(Sir Tom Blundell's FRS alumini)

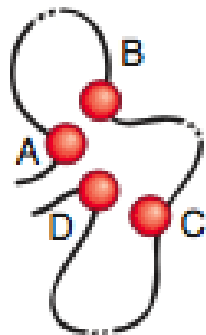


# Modern protein theory: two wrongs make one right

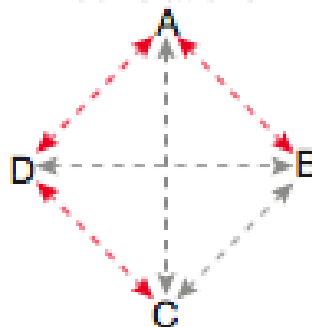
## Co-variance “plus” amino acid mutation modelling

- E.g. Burger and Nimwegen approaches
  - Techniques to model protein structures from co-evolving mutation frequencies (co-variance)
  - If co-evolving amino acids are identified mutating residue destabilises the protein structure and shifts its antigenicity, BUT mutating the second residue recovers the structure (and the antigenicity)

Physical contacts



Observed correlations



Predicted contacts

	A	B	C	D
A		■	■	■
B	■		■	
C				■
D	■		■	

■ Causative ■ Transitive

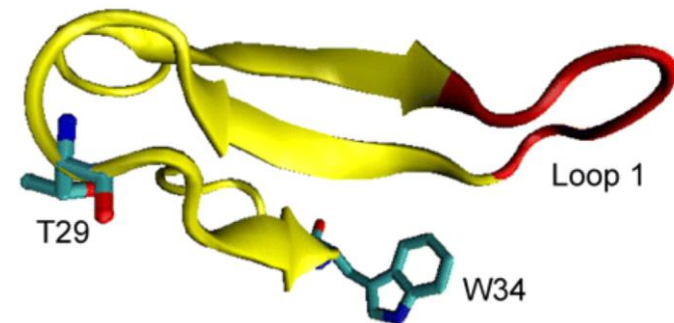
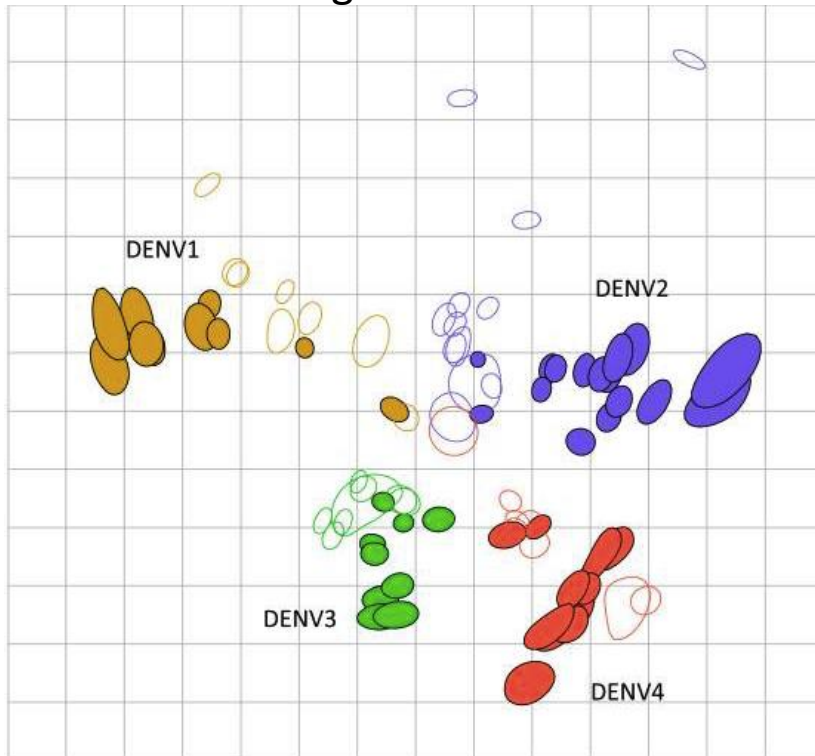


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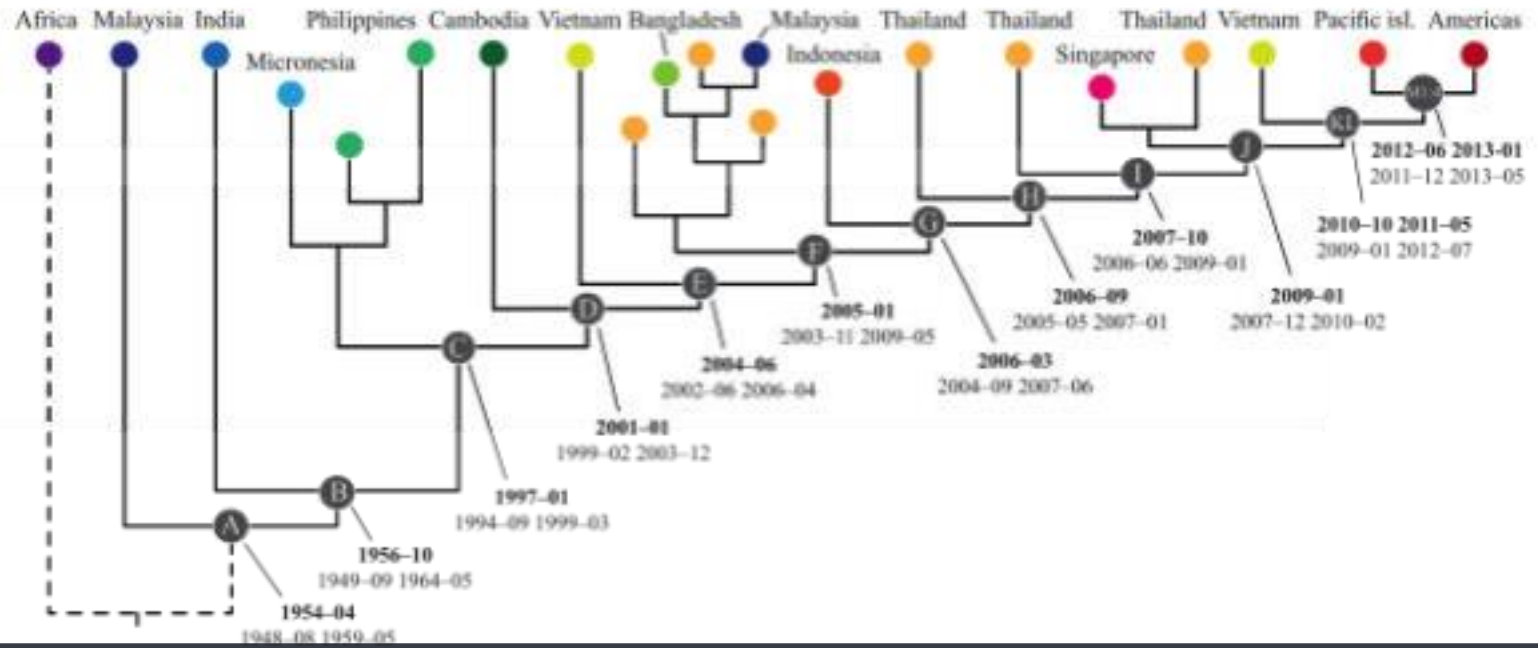
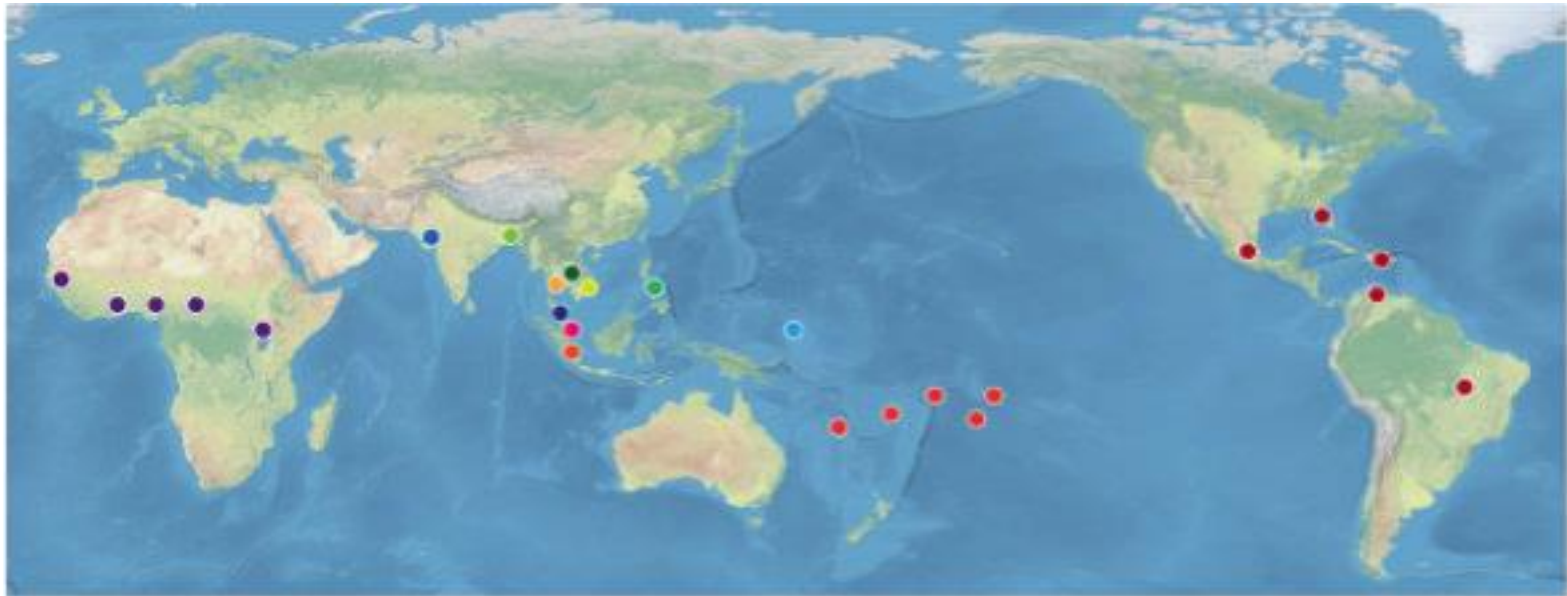
The state of the art protein modelling methods identify a loop (10 aa) in N-term. E protein showing clear co-evolution of residues and associated with ADE

prM and NS1 have remained elusive to this approach

Tested using infectious clones

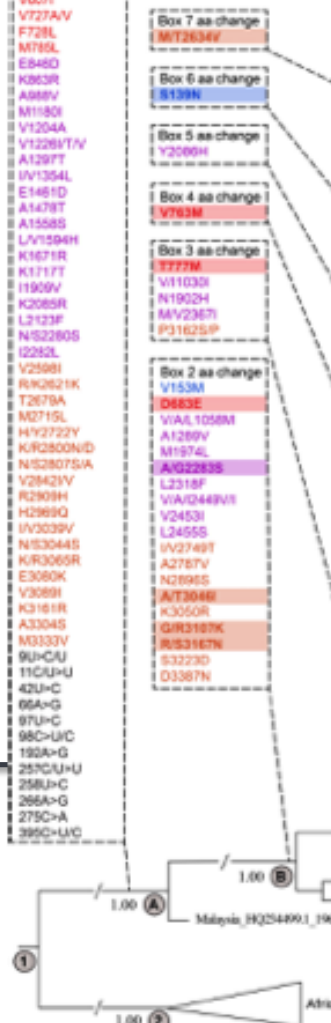


# Zika virus evolution: Africa to SE Asia

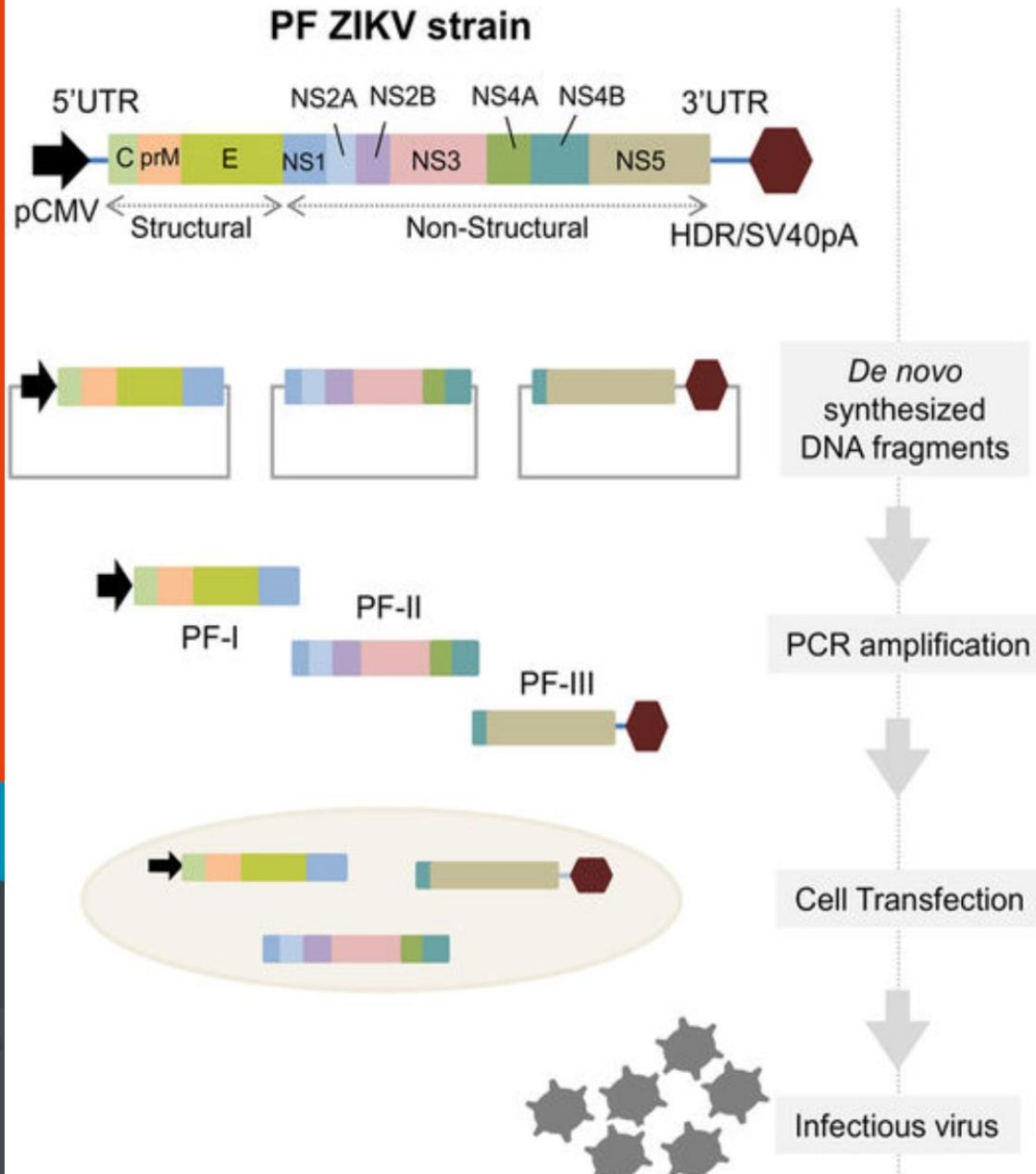


# Zika virus evolution: Africa to SE Asia

Box 1 aa and UTR change	Colour code box:	5'UTR (rRNA)
30A→U	Capid	1-122
35→U (indel)	PrM	123-200
S2A→G/A	Envelope	291-794
S8Q→A	NS1	795-1146
N258	NS2A	1147-1372
L27P	NS2B	1373-1502
R101K	NS3	1503-2119
I15V	NS4A	2120-2246
I125V	NS4B	2247-2520
K143E	NS5	2521-3423
A/V148P	3'UTR (rRNA)	
H157Y		
V158I		
K248R		
V/A295A		
V293A		
T410A		
V456I		
F/S475S		
V607I		
V727A/V		
F728L		
M795L		
E848D		
K853R		
A988V		
M1180I		
V1204A		
V1228V/T/V		
A1297T		
I/V1354L		
E1461D		
A1478T		
A1558S		
L/V1594H		
K1671R		
K1717T		
I1909V		
K2085R		
L2123P		
N/S2280S		
I2252L		
V2598I		
R/K2621K		
T2679A		
M2715L		
H/Y2722Y		
K/R2800N/D		
N/S2807S/A		
V2842V		
R2909H		
E2989Q		
I/V3039V		
N/S3044S		
K/R3065R		
E3080K		
V3089I		
K3181R		
A3304S		
M3337V		
9U→CAU		
T1CAU→U		
42U→C		
66A→G		
97U→C		
98C→U/C		
192A→G		
257CAU→U		
258U→C		
266A→G		
275C→A		
395Q→U/C		



# Zika virus reverse genetics ISA



Gibson assembly



# Acknowledgements

- 📍 Ernie Gould, Centre of Ecology, Oxford, UK
- 📍 Annelies Wilder-Smith – robust project management
- 📍 Duane Gubler, Dukes, USA
- 📍 Kevin Arien, ITM, Belgium (ZikaPLAN)
- 📍 Phil Minor, Mark Page, James Ashall, NIBSC, UK
- 📍 Suzanna Kaptein – help with mouse model
- 📍 Ludmila Lobkowicz, Nick Furnham, Tapan Bhattacharyya, LSHTM